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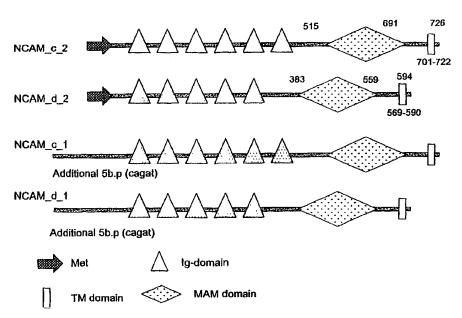
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(54) Title: N-CAM HOMOLOGS



(57) Abstract: The invention concerns several nucleic acid and amino acid sequences which are homologs of the neuronal cell adhesion molecule. These sequences are splice variants of each other.



For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

N-CAM HOMOLOGS

FIELD OF THE INVENTION

The present invention concerns novel nucleic acid sequences, vectors and host cells containing them, amino acid sequences encoded by said sequences, and antibodies reactive with said amino acid sequences, as well as pharmaceutical 5 compositions comprising any of the above. The present invention further concerns methods for screening for candidate activator or deactivators utilizing said amino acid sequences, as well as therapeutical and diagnostic utilization of said novel sequences.

BACKGROUND OF THE INVENTION

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Cell adhesion molecules (CAMs) are transmembrane glycoproteins with extracellular binding domains and cytoplasmic functional domains. Ligand binding to the extracellular domain initiates intracellular events through the cytoplastmic functional domain. These in turn cause a major behavioral and functional change in the cells. For cells to interact with each other or with their matrix, two 15 complementary molecules are required, the adhesion molecule and its ligand, one on each side of the adhesion site.

The immunoglobulin-like super family of adhesion molecules is a large and diverse family of molecules so named because they have one or more immunoglobulin-like domains. Included within the group are molecules concerned with antigen recognition by an adhesion to lymphocytes. These include CD3, CD4 and CD8 which together recognize complexes of antigen peptide and the major histocompatibility complex on other cells, and lymphocyte function related antigens such as CD2 and others of the LFA group of molecules.

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Other important molecular subgroups in this super family are the intercellular adhesion molecules (ICAM) which are widely expressed on epithelial and endothelial cells, nervous system adhesion molecules, e.g. neural cell adhesion molecules (NICAM), and molecules including L1 and TAG involved in organization and function of nerves.

Cell adhesion molecules are responsible for more than just adhesion of cells to one another and to their insoluble matrices. Additional functions include the ordering of cell sorting, migration and differentiation; organization of cell motility via the cytoskeleton; regulation of inter and intracellular signaling; and control gene transcription.

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Cell adhesion molecules (CAMs) are a subset of the immunoglobulin super family found in the nervous system of both vertebrates and invertebrates. They are usually surface membrane proteins with multiple Ig domains in their N-termini followed by several fibronectin-type III repeats and either a transmembrane intracellular domain or a glycophosphatidyl inositol-linked membrane anchor at the C-terminus.

The neuronal cell adhesion molecule (NCAM) is a member of the immunoglobulin super family and is strongly expressed in the nervous system. CAM is found in three major forms of which two – NCAM-140 and NCAM-180 are transmembrane proteins, while the third – NCAM-120 is attached to the membrane via a glycosylphosphatidyl inositol anchor. In addition, soluble NCAM forms exist in the brain, cerebrospinal fluid and in the plasma. NCAM mediates cell adhesion through homophilic as well as heterophilic activities. Following NCAM binding, transmembrane signalling is believed to be achieved resulting in increased intracellular calcium.

By mediating cell adhesion to other cells and to the extracellular matrix, and by activating intracellular signal pathways, NCAM influences cell migration, neurite extension and possibly also formation of synapses in the brain. From some studies on NCAM knock-out mice, it has been deduced that NCAM is crucial for the formation of the olfactory bulb and the mossy fiber system in the hippocampus.

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In addition, NCAM is important for neuronal plasticity in the adult brain associated with learning and regeneration.

Several mutations in the L1 CAM are associated with the wide spectrum of neurologic abnormalities and mental retardation. This spectrum includes X-linked hydrocephalus, MASA syndrome, X-linked complicated spastic paraplegia type 1 and X-linked agenesis of the corpus callosum. These four diseases were initially described as distinct clinical entities with an overlapping clinical spectrum, but can now be lumped into one syndrome caused by mutations in the L1 gene. The main clinical features of this spectrum are Corpus callosum hypoplasia, mental retardation, adducted thumbs, spastic paraplegia and hydrocephalus, which has led to the acronym CRASH syndrome.

This protein may be a secreted adhesion-like molecule with anti-protease activity. A mutated or partially deleted sequence results in Kallmann Syndrome which is a genetic disorder that associates hypogonadotropic hypogonadism and anosmia. In this disease, the normal embryonic migration of GNRH-synthesizing neurons from the olfactory placodes to the hypothalamic region as well as the axonal extension of olfactory neurons towards the forebrain are impaired.

It has been found that NCAM is expressed also in a number of different tissues and cell types, beyond neurons, such as muscles and endocrine-originating tumors, and can be detected in sera of patients with small-cell lung cancer.

In tumor, the role of NCAM is attributed to tumor invasion and formation of metastatis. Malignant plasma cells and a subset of plasma cells from patients with monoclonal gammopathy exhibit surface expression of NCAM whereas normal plasma cells do not express NCAM.

An isoform of NCAM which is rich in polysialic acids and characteristic for embryonal NCAM (eNCAM) has been shown to be elevated in sera of patients with multiple myeloma using a chemiluminescence immunoassay.

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GLOSSARY

In the following description and claims use will be made, at times, with a variety of terms, and the meaning of such terms as they should be construed in accordance with the invention is as follows:

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"Cell adhesion molecules homolog (CAM-H) nucleic acid sequence" - the sequence shown in SEQ ID NO: 1 or 7, sequences having at least 70% identity to said sequence and fragments of the above sequences being 20 b.p. long. These sequences contain an N-terminal which has a homology to neural cell adhesion 10 molecules containing several repeats of the Ig-like C2-domain which are immunoglobulin-like domains involved in protein-protein and protein-ligand interactions. In particular, the N-terminal part of the novel sequences of the invention are homologues to Contactin/F3-subgroup adhesion molecules; NB-2, Tag-1 (Axonin-1), Big-1, Big-2, and Contactin.

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The novel sequences of the invention contain in their C-terminal neuropilinlike MAM domain. Neuropilin is a calcium-independent cell adhesion molecule that functions during the formation of certain neuronal circuits. This protein binds to Semaphorin III and to the VEGF165 isoform of VEGF. The MAM domain has been recognized as the extracellular region of functionally diverse proteins. All the 20 protein have a modular receptor-like architecture consisting of a signal peptide, followed by a large N-terminal extracellular domain, a single transmembrane region and an intracellular region.

SEQ ID NOS: 1 to 6 are from Homo sapiens and SEQ ID NO: 7 is from mouse.

SEQ ID NO: 4 is in fact identical to SEQ ID NO: 1 - and was inserted under a different name for convenience reasons. SEQ ID NO: 5 is an update of SEQ ID NO: 2 (i.e. a sequence where 3 nucleotides have been altered).

SEQ ID NO: 1 (and 4), 2 (and 5), 3 and 6 are in fact all splice variants of each other.

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The term CAM-H does not necessarily signify that the protein coded by the above sequences has the same or even similar physiological activities to known

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CAM molecules and merely indicates that it shows sequence homology with two CAMs.

"CAM homology product (CAM-H product) – also referred at times as the "CAM-H protein" or "CAM-H polypeptide" – is an amino acid coded by the nucleic acid sequences of SEQ ID NO: 1 to SEQ ID NO: 7. The amino acid sequence may be a peptide, a protein, as well as peptides or proteins having chemically modified amino acids (see below) such as a glycopeptide or glycoprotein. An example of an CAM-H product is shown in SEQ ID NO: 8 to SEQ ID NO: 13 (coded by the amino acid sequences of SEQ ID NO: 1 to SEQ ID NO: 7 respectively). The term also includes analogues of said sequences in which one or more amino acids has been added, deleted, substituted (see below) or chemically modified (see below) as well as fragments of this sequence having at least 10 amino acids.

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"Nucleic acid sequence" – a sequence composed of DNA nucleotides, RNA nucleotides or a combination of both types and may includes natural nucleotides, chemically modified nucleotides and synthetic nucleotides.

"Amino acid sequence" – a sequence composed of any one of the 20 naturally appearing amino acids, amino acids which have been chemically modified (see below), or composed of synthetic amino acids.

"Fragment of CAM-H product" - a polypeptide which has an amino acid sequence which is the same as part of but not all of the amino acid sequence of the CAM-H product.

"Fragments of CAM-H nucleic acid sequence" a continuous portion, preferably of about 20 nucleic acid sequences of the CAM-H nucleic acid sequence.

"Conservative substitution" - refers to the substitution of an amino acid in one class by an amino acid of the same class, where a class is defined by common physicochemical amino acid side chain properties and high substitution frequencies in homologous proteins found in nature, as determined, for example, by a standard Dayhoff frequency exchange matrix or BLOSUM matrix. [Six general classes of amino acid side chains have been categorized and include: Class I (Cys); Class II (Ser, Thr, Pro, Ala, Gly); Class III (Asn, Asp, Gln, Glu); Class IV (His, Arg, Lys); Class V (Ile, Leu, Val, Met); and Class VI (Phe, Tyr, Trp). For example, substitution of an Asp for another class III residue such as Asn, Gln, or Glu, is a conservative substitution.

"Non-conservative substitution" - refers to the substitution of an amino acid in one class with an amino acid from another class; for example, substitution of an Ala, a class II residue, with a class III residue such as Asp, Asn, Glu, or Gln.

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"Chemically modified" - when referring to the product of the invention, means a product (protein) where at least one of its amino acid resides is modified either by natural processes, such as processing or other post-translational modifications, or by chemical modification techniques which are well known in the art. Among the numerous known modifications typical, but not exclusive examples include: acetylation, acylation, amidation, ADP-ribosylation, glycosylation, GPI anchor formation, covalent attachment of a lipid or lipid derivative, methylation, myristlyation, pegylation, prenylation, phosphorylation, ubiqutination, or any similar process.

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"Biologically active" - refers to the CAM-H product which has physiological, regulatory or biochemical functions on the same target sites which naturally occurring CAM-H effects, for example can bind to the same ligands as the CAM, for example ligands present in extracellular matrixes; can interact with the same ligands present on the surface cells with which CAMs interact etc.

"Immunologically active" defines the capability of a natural, recombinant or synthetic CAM-H product, or any fragment thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

Thus, for example, a biologically active fragment of CAM-H product denotes a fragment which retains some or all of the immunological properties of the CAM-H product, e.g can bind specific anti-CAM-H product antibodies or which can elicit an immune response which will generate such antibodies or cause proliferation of specific immune cells which produce CAM-H.

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"Optimal alignment" - is defined as an alignment giving the highest percent identity score. Such alignment can be performed using a variety of commercially available sequence analysis programs, such as the local alignment program LALIGN using a ktup of 1, default parameters and the default PAM. A preferred alignment is the one performed using the CLUSTAL-W program from MacVector (TM), operated with an open gap penalty of 10.0, an extended gap penalty of 0.1, and a BLOSUM similarity matrix. If a gap needs to be inserted into a first sequence to optimally align it with a second sequence, the percent identity is calculated using only the residues that are paired with a corresponding amino acid residue (i.e., the calculation does not consider residues in the second sequences that are in the "gap" of the first sequence).

"Having at least X% identity" - with respect to two amino acid or nucleic acid sequence sequences, refers to the percentage of residues that are identical in the two sequences when the sequences are optimally aligned. Thus, 70% amino acid sequence identity means that 70% of the amino acids in two or more optimally aligned polypeptide sequences are identical.

"Isolated nucleic acid molecule having an CAM-H nucleic acid sequence" - is a nucleic acid molecule that includes the coding CAM-H nucleic acid sequence.

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Said isolated nucleic acid molecule may include the CAM-H nucleic acid sequence as an independent insert; may include the CAM-H nucleic acid sequence fused to an additional coding sequences, encoding together a fusion protein in which the CAM-H coding sequence is the dominant coding sequence (for example, the additional coding sequence may code for a signal peptide); the CAM-H nucleic acid sequence may be in combination with non-coding sequences, e.g., introns or control elements, such as promoter and terminator elements or 5' and/or 3' untranslated regions, effective for expression of the coding sequence in a suitable host; or may be a vector in which the CAM-H protein coding sequence is a heterologous.

"Expression vector" - refers to vectors that have the ability to incorporate and express heterologous DNA fragments in a foreign cell. Many prokaryotic and eukaryotic expression vectors are known and/or commercially available.

15 Selection of appropriate expression vectors is within the knowledge of those having skill in the art.

"Deletion" - is a change in either nucleotide or amino acid sequence in which one or more nucleotides or amino acid residues, respectively, are absent.

"Insertion" or "addition" - is that change in a nucleotide or amino acid sequence which has resulted in the addition of one or more nucleotides or amino acid residues, respectively, as compared to the naturally occurring sequence.

"Substitution" - replacement of one or more nucleotides or amino acids by different nucleotides or amino acids, respectively. As regards amino acid sequences the substitution may be conservative or non-conservative.

"Antibody" - refers to IgG, IgM, IgD, IgA, and IgG antibody. The definition includes polyclonal antibodies or monoclonal antibodies. This term refers to

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whole antibodies or fragments of the antibodies comprising the antigen-binding domain of the anti-CAM-H product antibodies, e.g. antibodies without the Fc portion, single chain antibodies, fragments consisting of essentially only the variable, antigen-binding domain of the antibody, etc. The term "antibody" may also refer to antibodies against nucleic acid sequences obtained by cDNA vaccination.

"Activator" - as used herein, refers to a molecule which mimics the effect of the natural CAM-H product or at times even increases or prolongs the duration of the biological activity of said product, as compared to that induced by the natural product. The mechanism may be by binding to the same moiety to which native CAM-H binds (for example on the extracellular matrix or to a protein present on the membrane of other cells) thus mimicking the activity of CAM-H; by prolonging the lifetime of the CAM-H, (for example by decrease of the rate of its degradation), by increasing the affinity of CAM-H to moieties which it binds (such as the extracellular moieties) etc. Activators may be polypeptides, nucleic acids, carbohydrates, lipids, or derivatives thereof, or any other molecules which can bind to and activate the CAM-H product.

"Deactivator" or ("Inhibitor") - refers to a molecule which modulates the activity of the CAM-H product in an opposite manner to that of the activator, by decreasing or shortening the duration of the biological activity of the CAM-H product. This may be done by blocking the binding of the CAM-H to the moiety for example to the ligand to which it binds by competitive or non-competitive inhibition, by causing rapid degradation of the CAM-H, etc. by inhibiting association of the CAM-H with the effectors which regulate its expression. Deactivators may be polypeptides, nucleic acids, carbohydrates, lipids, or derivatives thereof, or any other molecules which bind to and modulate the activity of said product.

"Treating a disease" - refers to administering a therapeutic substance effective to ameliorate symptoms associated with a disease, to lessen the severity or cure the disease, or to prevent the disease from occurring.

"Detection" – refers to a method of detection of a disease. This term may refer to detection of a predisposition to a disease.

"Probe" - the CAM-H nucleic acid sequence, or a sequence complementary therewith, when used to detect presence of other similar sequences in a sample.

The detection is carried out by identification of hybridization complexes between the probe and the assayed sequence. The probe may be attached to a solid support or to a detectable label.

SUMMARY OF THE INVENTION

The present invention is based on the finding of novel cell adhesion molecules containing an N-terminal which has Ig-like C2 type repeats, and a C-terminal which contains a neuropilin-like MAM domain.

Thus the present invention provides by its first aspect, a novel isolated nucleic acid molecule comprising or consisting of the sequence of SEQ ID NO: 1 to SEQ ID NO: 7, fragments of said sequence having at least 20 nucleic acids, or a molecule comprising a sequence having at least 70%, preferably 80%, and most preferably 90% identity to SEQ ID NO:1 to SEQ ID NO: 7.

The present invention further provides a protein or polypeptide comprising or consisting of an amino acid sequence encoded by any of the above nucleic acid sequences, termed herein "CAM-H product", for example, an amino acid sequence having the sequence as depicted in SEQ ID NO: 8 to 13, fragments of the above amino acid sequence having a length of at least 10 amino acids, as well as homologs of the amino acid sequences of SEQ ID NO: 8 to 13 in which one or more of the amino acid residues has been substituted (by conservative or non-conservative substitution) added, deleted, or chemically modified.

The present invention further provides nucleic acid molecule comprising or consisting of a sequence which encodes the above amino acid sequences, (including the fragments and analogs of the amino acid sequences). Due to the degenerative nature of the genetic code, a plurality of alternative nucleic acid 5 sequences, beyond those depicted in SEQ ID NO:1 to SEQ ID NO:7, can code for the amino acid sequence of the invention. Those alternative nucleic acid sequences which code for the same amino acid sequences codes by the sequences of SEQ ID NO: 1 to SEQ ID NO: 7 are also an aspect of the of the present invention.

The present invention further provides expression vectors and cloning 10 vectors comprising any of the above nucleic acid sequences, as well as host cells transfected by said vectors.

The present invention still further provides pharmaceutical compositions comprising, as an active ingredient, said nucleic acid molecules, said expression vectors, or said protein or polypeptide.

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These pharmaceutical compositions are suitable for the treatment of diseases and pathological conditions, which can be ameliorated, cured or prevented by raising the level of the CAM-H product. Typically these are diseases which are manifested by non-normal levels of various CAMs, which are usually lower than normal levels of CAM, or alternatively, diseases in which the level of CAM is 20 normal, but a therapeutically beneficial effect may be achieved by raising the level of CAM to a higher than normal level. Usually, these diseases are concerned with interaction of cells at the extracellular matrix, or interaction with other cells which can lead to a plurality of diseases and pathological conditions, which are specified hereinbelow.

By this preferred embodiment the pharmaceutical compositions of the invention may stimulate growth and regeneration of nerve cell axons in order to compensate for defects caused by genetic, inflammatory, neurogenerative or trauma causes, for example to target neurite regrowth to target muscles and nerve cells after nerve injury.

In addition, the composition may be used for the treatment of a plurality of CAM-involved diseases such as: inflammatory diseases, autoimmune diseases (Crohn's disease, colitis, rheumatoid arthritis), graft vs. host and host vs. graft diseases, for the treatment of multiple sclerosis, diabetes, atherosclerosis, various types of cancer, for the treatment of injuries caused by head trauma as well as for the treatment of various viral disease and the treatment of various respiratory diseases.

By a second aspect, the present invention provides a nucleic acid molecule comprising or consisting of a non-coding sequence which is complementary to that of SEQ ID NO: 1 to SEQ ID NO: 7, or complementary to a sequence having at least 70% identity to said sequence or a fragment of said two sequences. The complementary sequence may be a DNA sequence which hybridizes with SEQ ID NO: 1 to SEQ ID NO: 7, or hybridizes to a portion of that sequence having a length sufficient to inhibit the transcription of the complementary sequence. The complementary sequence may be a DNA sequence which can be transcribed into an mRNA being an antisense to the mRNA transcribed from SEQ ID NO: 1 to SEQ ID NO: 7 or into an mRNA which is an antisense to a fragment of the mRNA transcribed from SEQ ID NO: 1 to SEQ ID NO: 7, so as to inhibit its translation. The complementary sequence may also be the mRNA or the fragment of the mRNA itself.

The nucleic acid sequences of the second aspect of the invention may be used for therapeutic or diagnostic applications for example for detection of the expression of CAM-H in various tissues, such as neuronal tissue, endothelial or epithelial tissue, tissues connected to the immune system, tissue obtained from tumors as well as body fluids such as cerebrospinal fluid, plasma and blood. Said detection may be indicative of a plurality of diseases, and pathological conditions stemming from genetic, inflammatory, infectious, degenerative or trauma causes. The diseases which can be detected are the same diseases mentioned above in connection with the treatment by the therapeutical compositions of the invention

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above. In addition, the detection may be indicative to the presence of tumors in general, and in particular to tumors from neuro-endocrine origin basis. In particular the level of any one of the CAM-H of the invention may be indicative of the invasiveness of the tumor indicating its stage of malignancy, which determination may be important for the prognosis of the patient, and for determining his/her best therapeutical modality.

The present invention also provides expression vectors comprising any one of the above defined complementary nucleic acid sequences and host cells transfected with said nucleic acid sequences or vectors, being complementary to those specified in the first aspect of the invention.

The invention also provides anti-CAM-H product antibodies, namely antibodies directed against the CAM-H product which specifically bind to said CAM-H product as well as antibodies which can be obtained through cDNA vaccination. Said antibodies are useful both for diagnostic and therapeutic purposes. For the diagnostic purposes they may be indicative of neuronal diseases and pathological conditions (such as for example inflammatory and autoimmune diseases, diseases of the respiratory and vascular tract, and indicative of tumors of neuro-endocrine origin and for the stage of malignancy as indicated above. In addition said antibody may be as an active ingredient in a pharmaceutical composition as will be explained below.

The present invention also provides pharmaceutical compositions comprising, as an active ingredient, the nucleic acid molecules which comprise or consist of said complementary sequences, or of a vector comprising said complementary sequences. The pharmaceutical composition thus provides pharmaceutical compositions comprising, as an active ingredient, said anti-CAM-H product antibodies.

The pharmaceutical compositions comprising said anti-CAM-H product antibodies or the nucleic acid molecule comprising said complementary sequence, are suitable for the treatment of diseases and pathological conditions where a therapeutically beneficial effect may be achieved by neutralizing the CAM-H or decreasing the amount of the CAM-H product or blocking its binding to its target

(for example the ligand to which it binds on cells), for example, by the neutralizing effect of the antibodies, or by the decrease of the effect of the antisense mRNA in decreasing expression level of the CAM-H product. Mostly these diseases are manifested by a higher than normal level of the CAM-H of the invention, or by 5 normal level of CAMs, however the disease may be ameliorated or a beneficial effect may be evident by decreasing said level. Examples of such diseases are for example tumors, and in particular of a neuro-endocrine origin, in which the invasiveness of the tumor is dependent on interactions between the CAM-H of the invention and basal membranes or extracellular matrixes, diseases involving the 10 immune system as well as endothelial and epithelial membranes, as well as other diseases of neural origin involving cell-adhesion molecules.

According to the third aspect of the invention the present invention provides methods for detecting the level of the transcript (mRNA) of said CAM-H product in a body fluid sample, plasma, cerebrospinal fluid, or in a specific tissue sample. 15 for example by use of probes comprising or consisting of said coding sequences; as well as methods for detecting levels of expression of said product in tissue, e.g. by the use of antibodies capable of specifically reacting with the above amino acid sequences. Detection of the level of the expression of the CAM-H of the invention may be indicative of a plurality of physiological or pathological conditions, as explained above.

The method, according to this latter aspect, for detection of a nucleic acid sequence which encodes the CAM-H product in a biological sample, comprises the steps of:

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- providing a probe comprising at least one of the nucleic acid (a) sequence defined above;
 - contacting the biological sample with said probe under conditions (b) allowing hybridization of nucleic acid sequences thereby enabling formation of hybridization complexes;
- detecting hybridization complexes, wherein the presence of the complex indicates the presence of nucleic acid sequence encoding the CAM-H product in the biological sample.

The amount of hybridization complexes may be determined and calibrated by comparing it to a calibration scale in order to determine the amount of the nucleic acid sequence which enables the CAM-H product in the sample. The level of each of the sequences SEQ ID NO: 1 to SEQ ID NO: 7 may be detected and either compared to each other, and said ratio may also be indicative to a plurality of pathological or physiological conditions as explained above.

By a preferred embodiment the probe is part of a nucleic acid chip used for detection purposes, i.e. the probe is a part of an array of probes each present in a known location on a solid support.

The nucleic acid sequence used in the above method may be a DNA sequence an RNA sequence, etc; it may be a coding or a sequence or a sequence complementary thereto (for respective detection of RNA transcripts or coding-DNA sequences). By quantization of the level of hybridization complexes and calibrating the quantified results it is possible also to detect the level of the transcript in the sample.

Methods for detecting mutations in the region coding for the CAM-H product are also provided, which may be methods carried-out in a binary fashion, namely merely detecting whether there is any mismatches between the normal CAM-H nucleic acid sequence and the one present in the sample, or carried-out by specifically detecting the nature and location of the mutation.

The present invention also concerns a method for detecting CAM-H product both for determining its presence, as well as its level or alterations in its level in a biological sample, comprising the steps of:

- (a) contacting with said biological sample the antibody of the invention, thereby forming an antibody-antigen complex; and
 - (b) detecting said antibody-antigen complex

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wherein the presence of said antibody-antigen complex correlates with the presence of CAM-H product in said biological sample.

The present invention also concerns a method for detecting anti-CAM-H antibodies in a biological sample comprising the steps of:

(a) contacting said biological sample with the product of the invention thereby forming an antibody-antigen complex; and

(b) detecting said antibody-antigen complex

wherein the presence of said antibody-antigen complex correlates with the presence of anti-CAM-H antibody in said biological sample.

In many cases, diseases are detected not by detecting the presence of the protein (product) which caused the disease, but rather by detecting the presence in a biological sample (such as blood or serum) of antibodies against such a product. The method of detecting the presence of anti-CAM-H antibodies is intended to be used in such case.

The amount of the antibody-antigen complex can be quantitized, in order to determine the level of the CAM-H-product or the anti-CAM-H antibodies, as the case may be.

As explained above the level of any of the products of SEQ ID NO: 8 (of the novel homolog) may be compared to each level, and the ratio between the levels may be indicative to a plurality of physiological and pathological conditions as explained above. In addition, the indicative ratio may not be the ratio of the proteins themselves but rather the ratio of antibodies against the proteins. Furthermore, the ratio of the level of the mRNA transcripts of any one of SQ ID NO: 1 to SEQ ID NO: 7, and changes in said ratio may be indicative of a plurality of diseases or pathological conditions especially as detailed above.

By yet another aspect the invention also provides a method for identifying candidate compounds capable of modulating the activity of CAM-H product (being either activators or deactivators). The method includes:

(i) providing a protein or polypeptide comprising an amino acid sequence substantially as depicted in SEQ ID NO: 8 to SEQ ID NO: 13, or a fragment of such a sequence;

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- (ii) contacting a candidate compound with said amino acid sequence;
- (iii) comprising the physiological effect of the amino acid sequence in the presence and absence of said candidate compound and selecting those compounds which show a significant effect on said physiological activity.

The activity of the amino acid which should be changed by the modulator (being either the activator or deactivator) may be for example the binding of the

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CAM-H product to a ligand of an adhesion molecule present for example on the external cell surface. Any modulator which changes such an activity has a potential as serving as an actuator or deactivator.

The present invention also concerns compounds identified by the above methods described above, which compound may either be an activator of the CAM-H product or a deactivator thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

In order to understand the invention and to see how it may be carried out in practice, a preferred embodiment will now be described, by way of non-limiting example only, with reference to the accompanying drawings, in which:

- Fig. 1 is the alignment of the CAM-H product of SEQ ID NO: 3 (NCAM_c_1), 4 (NCAM_c_2); 5 (NCAM_d_2) and 6 (NCAM_d_1);
- Fig. 2 is an alignment of human (SEQ ID NO: 4) and mouse (SEQ ID NO: 7) sequences;
 - Fig. 3 is an alignment of amino acid sequences of SEQ ID NO: 8 and 9;
 - Fig. 4 is an alignment of SEQ ID NOS: 10, 11, 12, 13;
 - Fig. 5 shows gel of PCR fragment of the sequence of the invention in various tissues; and
- Fig. 6 shows a schematic drawing of the amino acid sequence of SEQ ID NOS; 10, 11, 12, 13, showing the positions of Tm, Ig, and MAM domains.

DETAILED DESCRIPTION OF A PREFERRED EMBODIMENT

Example I: CAM-H - nucleic acid sequence

The nucleic acid sequences of the invention include nucleic acid sequences which encode CAM-H product and fragments and analogs thereof. The nucleic acid sequences may alternatively be sequences complementary to the above coding sequence, or to a region of said coding sequence. The length of the complementary sequence is sufficient to avoid the expression of the coding

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sequence. The nucleic acid sequences may be in the form of RNA or in the form of DNA, and include messenger RNA, synthetic RNA and DNA, cDNA, and genomic DNA. The DNA may be double-stranded or single-stranded, and if single-stranded may be the coding strand or the non-coding (anti-sense, 5 complementary) strand. The nucleic acid sequences may also both include dNTPs, rNTPs as well as non naturally occurring sequences. The sequence may also be a part of a hybrid between an amino acid sequence and a nucleic acid sequence.

In a general embodiment, the nucleic acid sequence has at least 70%, 10 preferably 80% or 90% sequence identity with the sequences identified as SEQ ID NO: 1 to SEQ ID NO: 7.

The nucleic acid sequences may include the coding sequence by itself. By another alternative the coding region may be in combination with additional coding sequences, such as those coding for fusion protein or signal peptides, in 15 combination with non-coding sequences, such as introns and control elements, promoter and terminator elements or 5' and/or 3' untranslated regions, effective for expression of the coding sequence in a suitable host, and/or in a vector or host environment in which the CAM-H nucleic acid sequence is introduced as a heterologous sequence.

The nucleic acid sequences of the present invention may also have the product coding sequence fused in-frame to a marker sequence which allows for purification of the CAM-H product. The marker sequence may be, for example, a hexahistidine tag to provide for purification of the mature polypeptide fused to the marker in the case of a bacterial host, or, the marker sequence may be a 25 hemagglutinin (HA) tag when a mammalian host, e.g. COS-7 cells, is used. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein (Wilson, I., et al. Cell 37:767 (1984)).

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Also included in the scope of the invention are fragments also referred to herein as oligonucleotides, typically having at least 20 bases, preferably 20-30 30 bases corresponding to a region of the coding-sequence nucleic acid sequence.

The fragments may be used as probes, primers, and when complementary also as antisense agents, and the like, according to known methods.

As indicated above, the nucleic acid sequence may be substantially a depicted in SEQ ID NO: 1 to SEO ID NO: 7 or fragments thereof or sequences 5 having at least 70%, preferably 70-80%, most preferably 90% identity to the above sequence. Alternatively, due to the degenerative nature of the genetic code. the sequence may be a sequence coding the amino acid sequence of SEQ ID NO: 8 to SEQ ID NO: 13, or fragments or analogs of said amino acid sequence.

Preparation of nucleic acid sequences. Α. 10

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The nucleic acid sequences may be obtained by screening cDNA libraries using oligonucleotide probes which can hybridize to or PCR-amplify nucleic acid sequences which encode the CAM-H products disclosed above. cDNA libraries prepared from a variety of tissues are commercially available and procedures for 15 screening and isolating cDNA clones are well-known to those of skill in the art. Such techniques are described in, for example, Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual (2nd Edition), Cold Spring Harbor Press, Plainview, N.Y. and Ausubel FM et al. (1989) Current Protocols in Molecular Biology, John Wiley & Sons, New York, N.Y.

The nucleic acid sequences may be extended to obtain upstream and downstream sequences such as promoters, regulatory elements, and 5' and 3' untranslated regions (UTRs). Extension of the available transcript sequence may be performed by numerous methods known to those of skill in the art, such as PCR or primer extension (Sambrook et al., supra), or by the RACE method using, for example, the Marathon RACE kit (Clontech, Cat. # K1802-1).

Alternatively, the technique of "restriction-site" PCR (Gobinda et al. PCR Methods Applic. 2:318-22, (1993)), which uses universal primers to retrieve flanking sequence adjacent a known locus, may be employed. First, genomic DNA is amplified in the presence of primer to a linker sequence and a primer specific to the known region. The amplified sequences are subjected to a second

round of PCR with the same linker primer and another specific primer internal to the first one. Products of each round of PCR are transcribed with an appropriate RNA polymerase and sequenced using reverse transcriptase.

Inverse PCR can be used to amplify or extend sequences using divergent primers based on a known region (Triglia, T. et al., Nucleic Acids Res. 16:8186, (1988)). The primers may be designed using OLIGO(R) 4.06 Primer Analysis Software (1992; National Biosciences Inc, Plymouth, Minn.), or another appropriate program, to be 22-30 nucleotides in length, to have a GC content of 50% or more, and to anneal to the target sequence at temperatures about 68-72°C.

The method uses several restriction enzymes to generate a suitable fragment in the known region of a gene. The fragment is then circularized by intramolecular ligation and used as a PCR template.

Capture PCR (Lagerstrom, M. et al., PCR Methods Applic. 1:111-19, (1991)) is a method for PCR amplification of DNA fragments adjacent to a known sequence in human and yeast artificial chromosome DNA. Capture PCR also requires multiple restriction enzyme digestions and ligations to place an engineered double-stranded sequence into a flanking part of the DNA molecule before PCR.

Another method which may be used to retrieve flanking sequences is that of Parker, J.D., et al., Nucleic Acids Res., 19:3055-60, (1991)). Additionally, one can use PCR, nested primers and PromoterFinderTM libraries to "walk in" genomic DNA (PromoterFinderTM; Clontech, Palo Alto, CA). This process avoids the need to screen libraries and is useful in finding intron/exon junctions. Preferred libraries for screening for full length cDNAs are ones that have been size-selected to include larger cDNAs. Also, random primed libraries are preferred in that they will contain more sequences which contain the 5' and upstream regions of genes.

A randomly primed library may be particularly useful if an oligo d(T) library does not yield a full-length cDNA. Genomic libraries are useful for extension into the 5' nontranslated regulatory region.

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The nucleic acid sequences and oligonucleotides of the invention can also be prepared by solid-phase methods, according to known synthetic methods. Typically, fragments of up to about 100 bases are individually synthesized, then joined to form continuous sequences up to several hundred bases.

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B. Use of CAM-H nucleic acid sequence for the production of CAM-H products

In accordance with the present invention, nucleic acid sequences specified above may be used as recombinant DNA molecules that direct the expression of CAM-H products.

As will be understood by those of skill in the art, it may be advantageous to produce CAM-H product-encoding nucleotide sequences possessing codons other than those which appear in SEQ ID NO: 1 to SEQ ID NO: 7 which are those which naturally occur in the human genome. Codons preferred by a particular prokaryotic or eukaryotic-Host (Murray, E. et al. Nuc Acids Res., 17:477-508, (1989)) can be selected, for example, to increase the rate of CAM-H product expression or to produce recombinant RNA transcripts having desirable properties, such as a longer half-life, than transcripts produced from naturally occurring sequence.

The nucleic acid sequences of the present invention can be engineered in order to alter a CAM-H product coding sequence for a variety of reasons, including but not limited to, alterations which modify the cloning, processing and/or expression of the product. For example, alterations may be introduced using techniques which are well known in the art, e.g., site-directed mutagenesis, to insert new restriction sites, to alter glycosylation patterns, to change codon preference, to produce splice variants, etc.

The present invention also includes recombinant constructs comprising one or more of the sequences as broadly described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a nucleic acid sequence of the invention has been inserted, in a forward or reverse orientation.

In a preferred aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and are commercially available. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are also described in Sambrook, et al., (supra).

The present invention also relates to host cells which are genetically engineered with vectors of the invention, and the production of the product of the invention by recombinant techniques. Host cells are genetically engineered (i.e., transduced, transformed or transfected) with the vectors of this invention which may be, for example, a cloning vector or an expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the expression of the CAM-H nucleic acid sequence. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to those skilled in the art.

The nucleic acid sequences of the present invention may be included in any one of a variety of expression vectors for expressing a product. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences, e.g., derivatives of SV40; bacterial plasmids; phage DNA; baculovirus; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. However, any other vector may be used as long as it is replicable and viable in the host.

The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease site(s) by procedures known in the art. Such procedures and related sub-cloning procedures are deemed to be within the scope of those skilled in the art.

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The DNA sequence in the expression vector is operatively linked to an appropriate transcription control sequence (promoter) to direct mRNA synthesis. Examples of such promoters include: LTR or SV40 promoter, the *E.coli lac* or trp promoter, the phage lambda PL promoter, and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. The expression vector also contains a ribosome binding site for translation initiation, and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. In addition, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E.coli*.

The vector containing the appropriate DNA sequence as described above, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. Examples of appropriate expression hosts include: bacterial cells, such as *E.coli*, *Streptomyces, Salmonella typhimurium*; fungal cells, such as yeast; insect cells such as *Drosophila* and *Spodoptera* Sf9; animal cells such as CHO, COS, HEK 293 or Bowes melanoma; adenoviruses; plant cells, etc. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein. The invention is not limited by the host cells employed.

In bacterial systems, a number of expression vectors may be selected depending upon the use intended for the CAM-H product. For example, when large quantities of CAM-H product are needed for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be desirable. Such vectors include, but are not limited to, multifunctional *E.coli* cloning and expression vectors such as *Bluescript*(R) (Stratagene), in which the CAM-H polypeptide coding sequence may be ligated into the vector in-frame with sequences for the amino-terminal Met and the subsequent 7 residues of beta-galactosidase so that a hybrid protein is produced;

pIN vectors (Van Heeke & Schuster J. Biol. Chem. 264:5503-5509, (1989)); pET vectors (Novagen, Madison WI); and the like.

In the yeast Saccharomyces cerevisiae a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase and 5 PGH may be used. For reviews, see Ausubel et al. (supra) and Grant et al., (Methods in Enzymology 153:516-544, (1987)).

In cases where plant expression vectors are used, the expression of a sequence encoding CAM-H product may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV (Brisson et al., Nature 310:511-514. (1984)) may be used alone or in combination with the omega leader sequence from TMV (Takamatsu et al., EMBO J., 6:307-311, (1987)). Alternatively, plant promoters such as the small subunit of RUBISCO (Coruzzi et al., EMBO J. 3:1671-1680, (1984); Broglie et al., Science 224:838-843, (1984)); or heat shock promoters (Winter J and Sinibaldi R.M., Results Probl. Cell Differ., 17:85-105, (1991)) may be used. These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. For reviews of such techniques, see Hobbs S. or Murry L.E. (1992) in McGraw Hill Yearbook of Science and Technology, McGraw Hill, New York, N.Y., pp 191-196; or Weissbach and Weissbach (1988) Methods for Plant Molecular Biology, Academic Press, New York, N.Y., pp 421-463.

CAM-H product may also be expressed in an insect system. In one such system, Autographa californica nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. The CAM-H product coding sequence may be cloned into a nonessential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of CAM-H coding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein coat. The recombinant viruses are then used to infect *S. frugiperda* cells or *Trichoplusia* larvae in which CAM-H protein is expressed (Smith *et al.*,

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J. Virol. 46:584, (1983); Engelhard, E.K. et al., Proc. Nat. Acad. Sci. <u>91</u>:3224-7, (1994)).

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, a CAM-H product coding sequence may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a nonessential E1 or E3 region of the viral genome will result in a viable virus capable of expressing CAM-H protein in infected host cells (Logan and Shenk, *Proc. Natl. Acad. Sci.* 81:3655-59, (1984). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Specific initiation signals may also be required for efficient translation of a CAM-H protein coding sequence. These signals include the ATG initiation codon and adjacent sequences. In cases where CAM-H product coding sequence, its initiation codon and upstream sequences are inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous transcriptional control signals including the ATG initiation codon must be provided. Furthermore, the initiation codon must be in the correct reading frame to ensure transcription of the entire insert. Exogenous transcriptional elements and initiation codons can be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate to the cell system in use (Scharf, D. et al., (1994) Results Probl. Cell Differ., 20:125-62, (1994); Bittner et al., Methods in Enzymol 153:516-544, (1987)).

In a further embodiment, the present invention relates to host cells containing the above-described constructs. The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell.

Introduction of the construct into the host cell can be effected by calcium

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phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis, L., Dibner, M., and Battey, I. (1986) Basic Methods in Molecular Biology). Cell-free translation systems can also be employed to produce polypeptides using RNAs derived from the DNA constructs of the present invention.

A host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the protein include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation and acylation. Post-translational processing which cleaves a "pre-pro" form of the protein may also be important for correct insertion, folding and/or function. Different host cells such as CHO, HeLa, MDCK, 293, WI38, etc. have specific cellular machinery and characteristic mechanisms for such post-translational activities and may be chosen to ensure the correct modification and processing of the introduced, foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express CAM-H product may be transformed using expression vectors which contain viral origins of replication or endogenous expression elements and a selectable marker gene.

Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clumps of stably transformed cells can be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler M., et al., Cell 11:223-32, (1977)) and adenine phosphoribosyltransferase (Lowy I., et al., Cell 22:817-23, (1980)) genes which can be employed in tk- or aprt- cells, respectively. Also, antimetabolite,

antibiotic or herbicide resistance can be used as the basis for selection; for example, *dhfr* which confers resistance to methotrexate (Wigler M., *et al.*, *Proc. Natl. Acad. Sci.* 77:3567-70, (1980)); *npt*, which confers resistance to the aminoglycosides neomycin and G-418 (Colbere-Garapin, F. *et al.*, *J. Mol. Biol.*, 5 150:1-14, (1981)) and *als* or *pat*, which confer resistance to CAM-Horsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been described, for example, *trpB*, which allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine (Hartman S.C. and R.C. Mulligan, *Proc. Natl. Acad. Sci.* 85:8047-51, (1988)). The use of visible markers has gained popularity with such markers as anthocyanins, beta-glucuronidase and its substrate, GUS, and luciferase and its substrates, luciferin and ATP, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C.A. *et. al.*, *Methods Mol. Biol.*, 55:121-131, (1995)).

Host cells transformed with a nucleotide sequence encoding CAM-H product may be cultured under conditions suitable for the expression and recovery of the encoded protein from cell culture. The product produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing nucleic acid sequences encoding CAM-H product can be designed with signal sequences which direct secretion of CAM-H product through a prokaryotic or eukaryotic cell membrane.

CAM-H product may also be expressed as a recombinant protein with one or more additional polypeptide domains added to facilitate protein purification. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle, Wash.). The inclusion of a

protease-cleavable polypeptide linker sequence between the purification domain and CAM-H protein is useful to facilitate purification. One such expression vector provides for expression of a fusion protein compromising a CAM-H polypeptide fused to a polyhistidine region separated by an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography, as described in Porath, et al., Protein Expression and Purification, 3:263-281, (1992)) while the enterokinase cleavage site provides a means for isolating CAM-H polypeptide from the fusion protein. pGEX vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to ligand-agarose beads (e.g., glutathione-agarose in the case of GST-fusions) followed by elution in the presence of free ligand.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period. Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification. Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents, or other methods, which are well know to those skilled in the art.

The CAM-H products can be recovered and purified from recombinant cell cultures by any of a number of methods well known in the art, including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography, and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the mature protein. Finally, high

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performance liquid chromatography (HPLC) can be employed for final purification steps.

C. Diagnostic applications utilizing nucleic acid sequences

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The nucleic acid sequences of the present invention may be used for a variety of diagnostic purposes. The nucleic acid sequences may be used to detect and quantitate expression of CAM-H in patient's cells, e.g. biopsied tissues, by detecting the presence of mRNA coding for CAM-H product. Alternatively, the assay may be used to detect soluble CAM-H in the serum or blood. This assay typically involves obtaining total mRNA from the tissue or serum and contacting the mRNA with a nucleic acid probe. The probe is a nucleic acid molecule of at least 20 nucleotides, preferably 20-30 nucleotides, capable of specifically hybridizing with a sequence included within the sequence of a nucleic acid molecule encoding CAM-H under hybridizing conditions, detecting the presence of mRNA hybridized to the probe, and thereby detecting the expression of CAM-H. This assay can be used to distinguish between absence, presence, and excess expression of CAM-H product and to monitor levels of CAM-H expression during therapeutic intervention.

The invention also contemplates the use of the nucleic acid sequences as a diagnostic for diseases resulting from inherited defective CAM-H sequences. These sequences can be detected by comparing the sequences of the defective (i.e., mutant) CAM-H coding region with that of a normal coding region. Association of the sequence coding for mutant CAM-H product with abnormal CAM-H product activity may be verified. In addition, sequences encoding 25 mutant CAM-H products can be inserted into a suitable vector for expression in a functional assay system (e.g., colorimetric assay, complementation experiments in a CAM-H protein deficient strain of HEK293 cells) as yet another means to verify or identify mutations. Once mutant genes have been identified, one can then screen populations of interest for carriers of the mutant gene.

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Individuals carrying mutations in the nucleic acid sequence of the present invention may be detected at the DNA level by a variety of techniques. Nucleic acids used for diagnosis may be obtained from a patient's cells, including but not limited to such as from blood, urine, saliva, placenta, tissue biopsy and autopsy material. Genomic DNA may be used directly for detection or may be amplified enzymatically by using PCR (Saiki, et al., Nature 324:163-166, (1986)) prior to analysis. RNA or cDNA may also be used for the same purpose. As an example, PCR primers complementary to the nucleic acid of the present invention can be used to identify and analyze mutations in the gene of the present invention.

Deletions and insertions can be detected by a change in size of the amplified product in comparison to the normal genotype.

Point mutations can be identified by hybridizing amplified DNA to radiolabeled RNA of the invention or alternatively, radiolabeled antisense DNA sequences of the invention. Sequence changes at specific locations may also be revealed by nuclease protection assays, such RNase and S1 protection or the chemical cleavage method (e.g. Cotton, et alProc. Natl. Acad. Sci. USA, 85:4397-4401, (1985)), or by differences in melting temperatures. "Molecular beacons" (Kostrikis L.G. et al., Science 279:1228-1229, (1998)), hairpin-shaped, single-stranded synthetic oligo- nucleotides containing probe sequences which are complementary to the nucleic acid of the present invention, may also be used to detect point mutations or other sequence changes as well as monitor expression levels of CAM-H product. Such diagnostics would be particularly useful for prenatal testing.

Another method for detecting mutations uses two DNA probes which are designed to hybridize to adjacent regions of a target, with abutting bases, where the region of known or suspected mutation(s) is at or near the abutting bases. The two probes may be joined at the abutting bases, e.g., in the presence of a ligase enzyme, but only if both probes are correctly base paired in the region of probe junction. The presence or absence of mutations is then detectable by the presence or absence of ligated probe.

Also suitable for detecting mutations in the CAM-H product coding sequence are oligonucleotide array methods based on sequencing by hybridization (SBH), as described, for example, in U.S. Patent No. 5,547,839. In a typical method, the DNA target analyte is hybridized with an array of oligonucleotides formed on a microchip. The sequence of the target can then be "read" from the pattern of target binding to the array.

D. Gene mapping utilizing nucleic acid sequences

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The nucleic acid sequences of the present invention are also valuable for chromosome identification. The sequence is specifically targeted to and can hybridize with a particular location on an individual human chromosome. Moreover, there is a current need for identifying particular sites on the chromosome. Few chromosome marking reagents based on actual sequence data (repeat polymorphisms) are presently available for marking chromosomal location. The mapping of DNAs to chromosomes according to the present invention is an important first step in correlating those sequences with genes associated with disease.

Briefly, sequences can be mapped to chromosomes by preparing PCR primers (preferably 20-30 bp) from the CAM-H cDNA. Computer analysis of the 3' untranslated region is used to rapidly select primers that do not span more than one exon in the genomic DNA, which would complicate the amplification process. These primers are then used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the primer will yield an amplified fragment.

PCR mapping of somatic cell hybrids or using instead radiation hybrids are rapid procedures for assigning a particular DNA to a particular chromosome. Using the present invention with the same oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes or pools of large genomic clones in an analogous manner. Other mapping strategies that can similarly be used to map to its chromosome include *in*

situ hybridization, prescreening with labeled flow-sorted chromosomes and preselection by hybridization to construct chromosome specific-cDNA libraries.

Fluorescence in situ hybridization (FISH) of a cDNA clone to a metaphase chromosomal spread can be used to provide a precise chromosomal location in one step. This technique can be used with cDNA as short as 50 or 60 bases. For a review of this technique, see Verma et al., Human Chromosomes: a Manual of Basic Techniques, (1988) Pergamon Press, New York.

Once a sequence has been mapped to a precise chromosomal location, the physical position of the sequence on the chromosome can be correlated with genetic map data. Such data are found, for example, in the OMIM database (Center for Medical Genetics, Johns Hopkins University, Baltimore, MD and National Center for Biotechnology Information, National Library of Medicine, Bethesda, MD). The OMIM gene map presents the cytogenetic map location of disease genes and other expressed genes. The OMIM database provides information on diseases associated with the chromosomal location. Such associations include the results of linkage analysis mapped to this interval, and the correlation of translocations and other chromosomal aberrations in this area with the advent of various diseases associated with abnormal amounts or function of various CAM proteins.

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E. Therapeutic applications of nucleic acid sequences

Nucleic acid sequences of the invention may also be used for therapeutic purposes. Turning first to the second aspect of the invention (i.e. inhibition of expression of CAM-H), expression of CAM-H product may be modulated through antisense technology, which controls gene expression through hybridization of complementary nucleic acid sequences, i.e. antisense DNA or RNA, to the control, 5' or regulatory regions of the gene encoding CAM-H product. For example, the 5' coding portion of the nucleic acid sequence sequence which codes for the product of the present invention is used to design an antisense oligonucleotide of from about 10 to 40 base pairs in length.

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Oligonucleotides derived from the transcription CAM-Ht site, e.g. between positions -10 and +10 from the CAM-Ht site, are preferred. An antisense DNA oligonucleotide is designed to be complementary to a region of the nucleic acid sequence involved in transcription (Lee et al., Nucl. Acids, Res., 6:3073, (1979); Cooney et al., Science 241:456, (1988); and Dervan et al., Science 251:1360, (1991)), thereby preventing transcription and the production of the CAM-H products. An antisense RNA oligonucleotide hybridizes to the mRNA in vivo and blocks translation of the mRNA molecule into the CAM-H products (Okano J. Neurochem. 56:560, (1991)). The antisense constructs can be delivered to cells by procedures known in the art such that the antisense RNA or DNA may be expressed in vivo. The antisense may be antisense mRNA or DNA sequence capable of coding such antisense mRNA. The antisense mRNA or the DNA coding thereof can be complementary to the full sequence of nucleic acid sequences coding to the CAM-H protein or to a fragment of such a sequence which is sufficient to inhibit production of a protein product.

Turning now to the first aspect of the invention, i.e. expression of CAM-H, expression of CAM-H product may be increased by providing coding sequences for coding for said product under the control of suitable control elements ending its expression in the desired host.

The nucleic acid sequences of the invention may be employed in combination with a suitable pharmaceutical carrier. Such compositions comprise a therapeutically effective amount of the compound, and a pharmaceutically acceptable carrier or excipient. Such a carrier includes but is not limited to saline, buffered saline, dextrose, water, glycerol, ethanol, and combinations thereof. The formulation should suit the mode of administration.

The products of the invention as well as any activators and deactivators compounds (see below) which are polypeptides, may also be employed in accordance with the present invention by expression of such polypeptides in vivo, which is often referred to as "gene therapy." Cells from a patient may be engineered with a nucleic acid sequence (DNA or RNA) encoding a polypeptide

ex vivo, with the engineered cells then being provided to a patient to be treated with the polypeptide. Such methods are well-known in the art. For example, cells may be engineered by procedures known in the art by use of a retroviral particle containing RNA encoding a polypeptide of the present invention.

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Similarly, cells may be engineered in vivo for expression of a polypeptide in vivo by procedures known in the art. As known in the art, a producer cell for producing a retroviral particle containing RNA encoding the polypeptide of the present invention may be administered to a patient for engineering cells in vivo and expression of the polypeptide in vivo. These and other methods for administering a product of the present invention by such method should be apparent to those skilled in the art from the teachings of the present invention. For example, the expression vehicle for engineering cells may be other than a retrovirus, for example, an adenovirus which may be used to engineer cells in vivo after combination with a suitable delivery vehicle.

Retroviruses from which the retroviral plasmid vectors mentioned above may be derived include, but are not limited to, Moloney Murine Leukemia Virus, spleen necrosis virus, retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, gibbon ape leukemia virus, human immunodeficiency virus, adenovirus, Myeloproliferative Sarcoma Virus, and mammary tumor virus.

The retroviral plasmid vector is employed to transduce packaging cell lines to form producer cell lines. Examples of packaging cells which may be transfected include, but are not limited to, the PE501, PA317, psi-2, psi-AM, PA12, T19-14X, VT-19-17-H2, psi-CRE, psi-CRIP, GP+E-86, GP+envAm12, and DAN cell lines as described in Miller (Human Gene Therapy, Vol. 1, pg. 25 5-14, (1990)). The vector may transduce the packaging cells through any means known in the art. Such means include, but are not limited to, electroporation, the use of liposomes, and CaPO₄ precipitation. In one alternative, the retroviral plasmid vector may be encapsulated into a liposome, or coupled to a lipid, and then administered to a host.

The producer cell line generates infectious retroviral vector particles which include the nucleic acid sequence(s) encoding the polypeptides. Such retroviral vector particles then may be employed, to transduce eukaryotic cells, either in vitro or in vivo. The transduced eukaryotic cells will express the nucleic acid sequence(s) encoding the polypeptide. Eukaryotic cells which may be transduced include, but are not limited to, embryonic stem cells, embryonic carcinoma cells, as well as hematopoietic stem cells, hepatocytes, fibroblasts, myoblasts, keratinocytes, endothelial cells, and bronchial epithelial cells.

The genes introduced into cells may be placed under the control of inducible promoters, such as the radiation-inducible Egr-1 promoter, (Maceri, H.J., et al., Cancer Res., 56(19):4311 (1996)), to stimulate CAM-H production or antisense inhibition in response to radiation, eg., radiation therapy for treating tumors.

5 Example II. CAM-H product

The substantially purified CAM-H product of the invention has been defined above as the product coded from the nucleic acid sequence of the invention. Preferably the amino acid sequence is an amino acid sequence having at least 70%, preferably at least 80% or 90% identity to the sequence identified as SEQ ID NO: 8 to SEQ ID NO: 13. The protein or polypeptide may be in mature and/or modified form, also as defined above. Also contemplated are protein fragments having at least 10 contiguous amino acid residues, preferably at least 10-20 residues, derived from the CAM-H product.

The sequence variations are preferably those that are considered conserved substitutions, as defined above. Thus, for example, a protein with a sequence having at least 80% sequence identity with the protein identified in SEQ ID NO: 8 to SEQ ID NO: 13, preferably by utilizing conserved substitutions as defined above is also part of the invention. In a more specific embodiment, the protein has or contains the sequence identified as SEQ ID NO: 8 to SEQ ID NO: 13. The CAM-H product may be (i) one in which one or more of the amino

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acid residues in a sequence listed above are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue), or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the CAM-H product is fused with another compound, 5 such as a compound to increase the half-life of the protein (for example, polyethylene glycol (PEG)), or a moiety which serves as targeting means to direct the protein to its target tissue or target cell population (such as an antibody), or (iv) one in which additional amino acids are fused to the CAM-H product. Such fragments, variants and derivatives are deemed to be within the scope of those skilled in the art from the teachings herein.

Preparation of CAM-H product A.

Recombinant methods for producing and isolating the CAM-H product, and fragments of the protein are described above.

In addition to recombinant production, fragments and portions of CAM-H product may be produced by direct peptide synthesis using solid-phase techniques (cf. Stewart et al., (1969) Solid-Phase Peptide Synthesis, WH Freeman Co, San Francisco; Merrifield J., J. Am. Chem. Soc., 85:2149-2154, (1963)). In vitro peptide synthesis may be performed using manual techniques or by automation. .20 Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer, Foster City, Calif.) in accordance with the instructions provided by the manufacturer. Fragments of CAM-H product may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

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B. Therapeutic uses and compositions utilizing the CAM-H product

The CAM-H product of the invention is generally useful in treating diseases and disorders which are characterized by a lower than normal level of CAM-H expression, and or diseases which can be cured or ameliorated by raising the level of the CAM-H product, even if the level is normal. Typically, there are

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diseases where non-normal growth, regeneration or tumors of neurons is evidenced due to genetic, degenerative or injury causes.

Typically these diseases are in CAM-H products or fragments and may be administered by any of a number of routes and methods designed to provide a consistent and predictable concentration of compound at the target organ or tissue. The product-containing compositions may be administered alone or in combination with other agents, such as stabilizing compounds, and/or in combination with other pharmaceutical agents such as drugs or hormones.

CAM-H product-containing compositions may be administered by a number of routes including, but not limited to oral, intravenous, intramuscular, transdermal, subcutaneous, topical, sublingual, or rectal means as well as by nasal application. CAM-H product-containing compositions may also be administered via liposomes. Such administration routes and appropriate formulations are generally known to those of skill in the art.

The product can be given via intravenous or intraperitoneal injection. Similarly, the product may be injected to other localized regions of the body. The product may also be administered via nasal insufflation. Enteral administration is also possible. For such administration, the product should be formulated into an appropriate capsule or elixir for oral administration, or into a suppository for rectal administration.

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The foregoing exemplary administration modes will likely require that the product be formulated into an appropriate carrier, including ointments, gels, suppositories. Appropriate formulations are well known to persons skilled in the art.

Dosage of the product will vary, depending upon the potency and therapeutic index of the particular polypeptide selected.

A therapeutic composition for use in the treatment method can include the product in a sterile injectable solution, the polypeptide in an oral delivery vehicle, the product in an aerosol suitable for nasal administration, or the product in a nebulized form, all prepared according to well known methods. Such

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compositions comprise a therapeutically effective amount of the compound, and a pharmaceutically acceptable carrier or excipient. Such a carrier includes but is not limited to saline, buffered saline, dextrose, water, glycerol, ethanol, and combinations thereof. The product of the invention may also be used to modulate 5 endothelial differentiation and proliferation as well as to modulate apoptosis either ex vivo or in vitro, for example, in cell cultures.

Example III. Screening methods for activators and deactivators (inhibitors)

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The present invention also includes an assay for identifying molecules, such as synthetic drugs, antibodies, peptides, or other molecules, which have a modulating effect on the activity of the CAM-H product, e.g. activators or deactivators of the CAM-H product of the present invention. Such an assay comprises the steps of providing an CAM-H product encoded by the nucleic acid 15 sequences of the present invention and determining its physiological activity on the target in the presence and absence of one or more candidate molecules to determine the candidate molecules. Those molecules which are modulating effect on the activity of the CAM-H product are selected as likely candidates for activators and deactivators.

CAM-H product, its catalytic or immunogenic fragments or oligopeptides thereof, can be used for screening therapeutic compounds in any of a variety of drug screening techniques. The fragment employed in such a test may be free in solution, affixed to a solid support, borne on a cell membrane or located intracellularly. The formation of binding complexes, between CAM-H product and the agent being tested, may be measured. Alternatively, the activator or deactivator may work by serving as agonist or antagonist, respectively, of the CAM-H receptor and their effect may be determined in connection with the receptor.

Another technique for drug screening which may be used provides for high throughput screening of compounds having suitable binding affinity to the CAM-H product is described in detail by Geysen in PCT Application WO

84/03564, published on Sep. 13, 1984. In summary, large numbers of different small peptide test compounds are synthesized on a solid substrate, such as plastic pins or some other surface. The peptide test compounds are reacted with the full CAM-H product or with fragments of CAM-H product and washed. Bound . 5 CAM-H product is then detected by methods well known in the art. Substantially purified CAM-H product can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

Antibodies to the CAM-H product, as described in Example IV below, may also be used in screening assays according to methods well known in the art. For example, a "sandwich" assay may be performed, in which an anti-CAM-H antibody is affixed to a solid surface such as a microtiter plate and CAM-H product is added. Such an assay can be used to capture compounds which bind to 15 the CAM-H product. Alternatively, such an assay may be used to measure the ability of compounds to influence with the binding of CAM-H product to the CAM-H receptor and then select those compounds which effect the binding.

Example IV. Anti-CAM-H antibodies

A. Synthesis 20

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In still another aspect of the invention, the purified CAM-H product is used to produce anti-CAM-H antibodies which have diagnostic and therapeutic uses related to the activity, distribution, and expression of the CAM-H product, in particular therapeutic applications in modulating the effect of CAM-H on moieties to which it binds in the extracellular matrix.

Antibodies to CAM-H product may be generated by methods well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, humanized, single chain, Fab fragments and fragments produced by an Fab expression library. Antibodies, i.e., those which inhibit dimer formation, are especially preferred for therapeutic use.

A fragment CAM-H product for antibody induction does not require biological activity but have to feature immunological activity; however, the protein fragment or oligopeptide must be antigenic. Peptides used to induce specific antibodies may have an amino acid sequence consisting of at least five amino acids, preferably at least 10 amino acids of the sequences specified in SEQ ID NO: 8 to SEQ ID NO: 13. Preferably they should mimic a portion of the amino acid sequence of the natural protein and may contain the entire amino acid sequence of a small, naturally occurring molecule. Short stretches of CAM-H protein amino acids may be fused with those of another protein such as keyhole limpet hemocyanin and antibody produced against the chimeric molecule. Procedures well known in the art can be used for the production of antibodies to CAM-H product.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, etc may be immunized by injection with CAM-H product or any portion, fragment or oligopeptide which retains immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include but are not limited to Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and dinitrophenol. BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are potentially useful human adjuvants.

Monoclonal antibodies to CAM-H protein may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include but are not limited to the hybridoma technique originally described by Koehler and Milstein (*Nature* 256:495-497, (1975)); the human B-cell hybridoma technique (Kosbor et al., Immunol. Today 4:72, (1983); Cote et al., Proc. Natl. Acad. Sci. 80:2026-2030, (1983)) and the EBV-hybridoma technique (Cole, et al., Mol. Cell Biol. 62:109-120, (1984)).

Techniques developed for the production of "chimeric antibodies", the splicing of mouse antibody genes to human antibody genes to obtain a molecule

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with appropriate antigen specificity and biological activity can also be used (Morrison et al., Proc. Natl. Acad. Sci. 81:6851-6855, (1984); Neuberger et al., Nature 312:604-608, (1984); Takeda et al., Nature 314:452-454, (1985)). Alternatively, techniques described for the production of single chain antibodies (U.S. Pat. No. 4,946,778) can be adapted to produce single-chain antibodies specific for the CAM-H protein.

Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening recombinant immunoglobulin libraries or panels of highly specific binding reagents as disclosed in Orlandi *et al.* (*Proc. Natl. Acad. Sci.* 86:3833-3837, 1989)), and Winter G and Milstein C., (*Nature* 349:293-299, (1991)).

Antibody fragments which contain specific binding sites for CAM-H protein may also be generated. For example, such fragments include, but are not limited to, the F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity (Huse W.D. et al., Science 256:1275-1281, (1989)).

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B. Diagnostic applications of antibodies

A variety of protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the formation of complexes between CAM-H product and its specific antibody and the measurement of complex formation. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two noninterfering epitopes on a specific CAM-H product is preferred, but a competitive binding assay may also be employed. These assays are described in Maddox D.E., et al.,

(J. Exp. Med. 158:1211, (1983)).

Antibodies which specifically bind CAM-H product are useful for the diagnosis of conditions or diseases characterized by over or under expression of CAM-H. Alternatively, such antibodies may be used in assays to monitor patients being treated with CAM-H product, its activators, or its deactivators. Diagnostic assays for CAM-H protein include methods utilizing the antibody and a label to detect CAM-H product in human body fluids or extracts of cells or tissues. The products and antibodies of the present invention may be used with or without modification. Frequently, the proteins and antibodies will be labeled by joining them, either covalently or noncovalently, with a reporter molecule. A wide variety of reporter molecules are known in the art.

A variety of protocols for measuring CAM-H product, using either polyclonal or monoclonal antibodies specific for the respective protein are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescent activated cell sorting (FACS). As 15 noted above, a two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on CAM-H product is preferred, but a competitive binding assay may be employed. These assays are described, among other places, in Maddox, et al. (supra). Such protocols provide a basis for diagnosing altered or abnormal levels of CAM-H product expression. Normal or standard values for CAM-H product expression are established by combining body or cell extracts taken from normal subjects, preferably human, with antibody to CAM-H product under conditions suitable for complex formation which are well known in the art. The amount of standard complex formation may be quantified by various methods, preferably by photometric 25 methods. Then, standard values obtained from normal samples may be compared with values obtained from samples from subjects potentially affected by disease. Deviation between standard and subject values establishes the presence of disease state.

The antibody assays are useful to determine the level of CAM-H present in a body fluid sample, in order to determine whether it is being overexpressed or

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underexpressed in the tissue, or as an indication of how CAM-H levels are responding to drug treatment.

Another alternative is to determine the presence and/or level of naturally occurring anti-CAM-H antibodies in a sample, such as blood or serum. Many times diseases are identified by detecting the presence or level of antibodies against a specific product. For the detection of such naturally occurring anti-CAM-H antibodies, the sample may be contacted with the product of the invention, for example as depicted in SEQ ID NO: 3 or SEQ ID NO: 4, or with an antigenic fragment thereof, and the presence or level of antibody-antigen complexes may be determined by methods well known in the art.

C. Therapeutic uses of antibodies

In addition to their diagnostic use the antibodies may have a therapeutical utility in blocking or decreasing the activity of the CAM-H product in pathological conditions where beneficial effect can be achieved by such a decrease.

The antibody employed is preferably a humanized monoclonal antibody, or a human Mab produced by known globulin-gene library methods. The antibody is administered typically as a sterile solution by IV injection, although other parenteral routes may be suitable. Typically, the antibody is administered in an amount between about 1-15 mg/kg body weight of the subject. Treatment is continued, e.g., with dosing every 1-7 days, until a therapeutic improvement is seen.

Although the invention has been described with reference to specific methods and embodiments, it is appreciated that various modifications and changes may be made without departing from the invention.

EXAMPLE V. Specific localizations of the sequence of the invention in brain tissue

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RT-PCR

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Five µg of total RNA were isolated from human tissues. The reaction was performed in a final volume of 20 µl and also contained Superscript II Reverse Transcriptase (Gibco/BRL, Gaithersburg, MD), 1xBuffer supplied by the manufacturer, 30 units of Rnasin (Promega, Medison, WI) and 10 pmol of oligoDT (Promega, Medison, WI).

Gene-specific primers were designed, antisense and sense PCR was carried out in a reaction mixture containing 1 μ l of 5xBuffer (Boehringer-Mannheim, Germany), 4 μ l of 2.5 mM dNTPs, 25 pmol of primers and 2.5 units Expand Long Template PCR System (Boehringer-Mannheim, Germany).

The cycling conditions were 94°C-3 min. Taq polymerase followed by 30 cycles of 94°C for 30 sec, 68°C-1 min and final extension at 68°C for 10 mins. The PCR reaction on PTC-225 (MJ Research, Inc).

The resulting sequences were separated on a gel and the results are shown in Fig. 5. As can be seen the transcript (from SEQ ID NO: 3) was identified in Brain Cerebellum and was not identified in tissues obtained from other tissues proving that the sequences of the invention are indeed unique to brain.

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CLAIMS:

- 1. An isolated nucleic acid sequence selected from the group consisting of:
- (i) the nucleic acid sequence depicted in SEQ ID NO: 1 to SEQ ID NO: 7;
- 5 (ii) nucleic acid sequences having at least 70% identity with the sequence of (i); and
 - (iii) fragments of (i) or (ii) of at least 20 b.p.
 - 2. A nucleic acid sequence according to Claim 1(ii) wherein the nucleic acid sequences have at least 80% identity with the sequence of Claim 1(i).
- 3. A nucleic acid sequence according to Claim 2, wherein the nucleic acid sequences have at least 90% identity.
 - 4. An isolated nucleic acid sequence complementary to the nucleic acid sequence of Claim 1.
 - 5. An amino acid sequence selected from the group consisting of:
- 15 (i) an amino acid sequence coded by the isolated nucleic acid sequence of Claim 1;
 - (ii) fragments of the amino acid squence of (i) having at least 10 amino acids;
 - (iii) analogues of the amino acid sequences of (i) or (ii) in which one or more amino acids has been added, deleted, replaced or chemically modified without substantially altering the biological activity of the parent amino acid sequence.
 - 6. An amino acid sequence according to Claim 5, as depicted in SEQ ID NO: 8 to SEQ ID NO: 13.
- 7. An isolated nucleic acid sequence coding for the amino acid sequence of Claim 5 or 6.
 - 8. A purified antibody which binds specifically to the amino acid sequence of Claim 5 or 6.

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9. An expression vector comprising the nucleic acid sequences of Claim 1 or 7 and control elements for the expression of the nucleic acid sequence in a suitable host.

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- 10. An expression vector comprising the nucleic acid sequence of Claim 4, and control elements for the expression of the nucleic acid sequence in a suitable host.
 - 11. A host cell transfected by the expression vector of Claim 9 or 10.
 - 12. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and as an active ingredient an agent selected from the group consisting of:
 - (i) the expression vector of Claim 9; and

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- (ii) the amino acid sequence of Claim 5 or 6.
- 13. A pharmaceutical composition according to Claim 12, for treatment of diseases which can be ameliorated, cured or prevented by raising the level of the Cell Adhesion Molecule Homolog (CAM-H).
- 14. A pharmaceutical composition comprising a pharmaceutically acceptable carrier and as an active ingredient an agent selected from the group consisting of:
 - (i) the nucleic acid sequence of Claim 4;
 - (ii) the expression vector of Claim 10; and
 - (iii) the purified antibody of Claim 8.
- 15. A pharmaceutical composition according to Claim 14, for treatment of diseases which can be ameliorated or cured by decreasing the level of the CAM-H product.
 - 16. A method for detecting an CAM-H nucleic acid sequence in a biological sample, comprising the steps of:
- (a) hybridizing to nucleic acid material of said biological sample a nucleic acid sequence of Claim 1 or 4; and
 - (b) detecting said hybridization complex;

wherein the presence of said hybridization complex correlates with the presence of an CAM-H nucleic acid sequence in the said biological sample.

17. A method according to Claim 16, wherein the nucleic acid material of said biological sample are mRNA transcripts.

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18. A method according to Claim 16, where the nucleic acid sequence is present in a nucleic acid chip.

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- 19. A method for identifying candidate compounds capable of binding to the CAM-H product and modulating its activity the method comprising:
- (i) providing a protein or polypeptide comprising an amino acid sequence substantially as depicted in SEQ ID NO: 8 to SEQ ID NO: 13, or a fragment of such a sequence;
- (ii) comparing the physiological effect of the CAM-H product in the absence and presence of said candidate compound and selecting those compounds
 which show a significant effect on said physiological activity.
 - 20. A method according to Claim 19, wherein the compound is an activator and the measured effect is increase in the physiological activity.
 - 21. A method according to Claim 19, wherein the compound is an deactivator and the effect is decrease in the physiological activity.
- 15 22. An activator of the amino acid sequence of Claim 5 or 6.
 - 23. An deactivator of the amino acid sequence of Claims 5 or 6.
 - 24. A method for detecting CAM-H-product in a biological sample, comprising the steps of:
- (a) contacting with said biological sample the antibody of Claim 8, thereby forming an antibody-antigen complex; and
 - (b) detecting said antibody-antigen complex

wherein the presence of said antibody-antigen complex correlates with the presence of CAM-H product in said biological sample.

- 25. A method for detecting anti-CAM-H antibodies in a biological sample comprising the steps of:
 - (a) contacting with said biological sample the antibody of Claim 5 or 6, thereby forming an antibody-antigen complex; and
 - (b) detecting said antibody-antigen complex

wherein the presence of said antibody-antigen complex correlates with the presence of anti-CAM-H antibody in said biological sample.

										••	••								
20	TCAGCTCCTC	TCAGCTCCTC	TCAGCTCCTC	TCAGCTCCTC	100	TGAAGAGGAG	TGAAGAGGAG	TGAAGAGGAG	TGAAGAGGAG	150	CCCTtGAGTT	CCCTTGAGTT	CCCTtGAGTT	CCCTtGAGTT	200	TGGaccaaaa	TGGaccaaaa	TGGacCAAAA	TGGacCAAAA
	ATGTCTTGCT	ATGTCTTGCT	ATGTCTTGCT	ATGTCTTGCT		CCTGTAACAT	CCTGTAACAT	CCTGTAACAT	CCTGTAACAT		GAAGGAGAAA	GAAGGAGAAA	gAAgGAGaAA	gAAgGAGaAA		AcagatCAGG	ACAGG	ACAGG	ATCCACGTCc AcagatCAGG
	AGGATCAGAG	AGGATCAGAG	AGGATCAGAG	AGGATCAGAG		TCAGGCTTGG	TCAGGCTTGG	TCAGGCTTGG	TCAGGCTTGG		CACTATCCGG	CACTATCCGG	CACTATCCGG	CACTATCcGG		ATCCACGTCC	ATCCACGTCC	ATCCACGTCC	
•	AACAAGGCAG	AACAAGGCAG	AACAAGGCAG	AACAAGGCAG		TATTGTGCAC	TATTGTGCAC	TATTGTGCAC	TATTGTGCAC		AAAGGGTCTA	AAAGGGTCTA	AAAGGGTCTA	AAAGGGTCTA		GTCACAGGaC	GTCACAGGaC	GTCACAGGaC	GTCACAGGaC
	GTTTTTACC	GTTTTTACC	GTTTTTACC	GTTTTTACC		CCACGGTTCG	CCACGGTTCG	CCACGGTTCG	CCACGGTTCG		CGCTACTCCG	CGCTACTCCG	CGCTACTCCG	CGCTACTCCG		GACTTGTCTA	GACTTGTCTA	GACTTGTCTA	GACTIGICTA
—	NCAM c 1	NCAM c 2	NCAM d 2	NCAM_d_1	51	NCAM c 1	NCAM C 2	NCAM d 2	NCAM d 1	101	NCAM c 1	NCAM C 2	NCAM_d_2	NCAM_d_1	151	NCAM c 1	NCAM c 2	NCAM d 2	NCAM d 1

250 CTTCAATGAG CTTCAATGAG CTTCAATGAG	300 GGTATTACTG GGTATTACTG GGTATTACTG	350 gataaagtca atcagagtgg gataaagtca atcagagtgg gataaagtca atcagagtgg
ACTCAAGTGT ACTCAAGTGT ACTCAAGTGT ACTCAAGTGT	300 TCAGCGACAC CAAGGAGGCC GGTATTACTG TCAGCGACAC CAAGGAGGCC GGTATTACTG TCAGCGACAC CAAGGAGGCC GGTATTACTG TCAGCGACAC CAAGGAGGCC GGTATTACTG	gataaagtca gataaagtca gataaagtca gataaagtca
TGCCTCTGAC AGATTCCAAG ACTCAAGTGT TGCCTCTGAC AGATTCCAAG ACTCAAGTGT TGCCTCTGAC AGATTCCAAG ACTCAAGTGT TGCCTCTGAC AGATTCCAAG ACTCAAGTGT	TCAGCGACAC TCAGCGACAC TCAGCGACAC	GGTCTCCAGC GGTCTCCAGC GGTCTCCAGC
TGCcTcTGaC TGCcTcTGaC TGCcTcTGaC	TTACAAATAT TTACAAATAT TTACAAATAT TTACAAATAT	AATGGCTTGG AATGGCTTGG AATGGCTTGG
CAGCAGGAAG CAGCAGGAAG CAGCAGGAAG CAGCAGGAAG	aCTTTGAGGA aCTTTGAGĠA aCTTTGAGGA aCTTTGAGGA	TAAAGCAGAG AATGGCTTGG C TAAAGCAGAG AATGGCTTGG C TAAAGCAGAG AATGGCTTGG C
201 NCAM_C_1 NCAM_C_2 NCAM_d_2 NCAM_d_1	251 NCAM_C_1 NCAM_C_2 NCAM_G_2 NCAM_G_2	301 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_1

CTGTTCATCA AAGTATAGGT CTGTTCATCA AAGTATAGGT CIGITCAICA AAGIATAGGI AAGTATAGGT CTGTTCATCA CCAGTAGTAA CCAGTAGTAA CCAGTAGTAA CCAGTAGTAA TTTGGATGAT TTTGGATGAT TTTGGATGAT TTTGGATGAT ATGTATACTA ATGTATACTA ATGTATACTA ATGTATACTA NCAM C 1 NCAM C 2 NCAM d 2 NCAM d 1 351

Fig. 1 (Cont. 1)

450	TCCGGTGTGT	TCCGGTGTGT	TCCGGTGTGT	TCCGGTGTGT	200	GGCCAGGAGG	GGCCAGGAGG	GGCCAGGAGG	GGCCAGGAGG	550	ACCATTCTTT	ACCATTCTTT	ACCATICITI	ACCATTCTTT	009	GACCTCAGGA	GACCTCAGGA	GACCTCAGGA	GACCTCAGGA
	ACAGTGTTCC	ACAGTGTTCC	ACAGTGTTCC	ACAGTGTTCC		CTGGAGACGT	CTGGAGACGT	CTGGAGACGT	CTGGAGACGT		AGATTTATGA	AGATTTATGA	AGATTTATGA	AGATTTATGA		AAGAATCTTC	AAGAATCTTC	AAGAATCTTC	AAGAATCTTC
	CTATGAGAGA	CTATGAGAGA	CTATGAGAGA	CTATGAGAGA		TTCGGTATAG	TTCGGTATAG	TTCGGTATAG	TTCGGTATAG		AAAGGAGTTG	AAAGGAGTTG	AAAGGAGTTG	AAAGGAGTTG		CTTAAAACTA	CTTAAAACTA	CTTAAAACTA	CTTAAAACTA
	AACAATTTTa	AACAATTTTa	AACAATTTTa	AACAATTTTa		AATCCTCCTG	AATCCTCCTG	AATCCTCCTG	AATCCTCCTG		AGGATCTGAT	AGGATCTGAT	AGGATCTGAT	AGGATCTGAT		AAACAAAGAT	AAACAAAGAT	AAACAAAGAT	AAACAAAGAT
	GAAGCTAAAg	GAAGCTAAAg	GAAGCTAAAg	GAAGCTAAAg		TGCCAATTCC	TGCCAATTCC	TGCCAATTCC	TGCCAATTCC		TCTTGCTGCA	TCTTGCTGCA	TCTTGCTGCA	TCTTGCTGCA		ACCCAGGGTG	ACCCAGGGTG	ACCCAGGGTG	ACCCAGGGTG
401				NCAM_d_1	451	NCAM c 1	NCAM C 2	NCAM d 2	NCAM_d_1	501	NCAM C 1	NCAM c 2	NCAM d 2	NCAM_d_1	551	NCAM c 1	NCAM C 2	NCAM_d_2	NCAM_d_1

Fig. 1(Cont. 2)

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650	TGTAATATTC	TGTAATATTC	TGTAATATTC	TGTAATATTC	700	ATCACCGTCA	ATCACCGTCA	ATCACCGTCA	ATCACCGTCA		750	GAGAGGCCAT	GAGAGGCCAT	GAGAGGCCAT	GAGAGGCCAT	0	900	TCTCTCACCT	TCTCTCACCT	TCTCTCACCT	TCTCTCACCT	
	GAGGAATGTA	GAGGAATGTA	GAGGAATGTA	GAGGAATGTA		ATAAAACAGC	ATAAAACAGC	ATAAAACAGC	ATAAAACAGC			GTAAATCCTG	GTAAATCCTG	GTAAATCCTG	GTAAATCCTG			GCCTGCACCT	GCCTGCACCT	GCCTGCACCT	GCCTGCACCT	
	TIGCTICAGI	TTGCTTCAGT	TTGCTTCAGT	TTGCTTCAGT		AGACTGTCCA	AGACTGTCCA	AGACTGTCCA	AGACTGTCCA			TCCTATAGTT	TCCTATAGTT	TCCTATAGTT	TCCTATAGTT			CAGGAGGAGA	CAGGAGGAGA	CAGGAGGAGA	CAGGAGGAGA	
	TATAGCTGCA	TATAGCTGCA	TATAGCTGCA	TATAGCTGCA		GGTGTCGTTT	GGTGTCGTTT	GGTGTCGTTT	GGTGTCGTTT			TGGTGGATGA	TGGTGGATGA	TGGTGGATGA	TGGTGGATGA			TGTGTTACAA	TGTGTTACAA	TGTGTTACAA	TGTGTTACAA	
	CTATGCTAAT	CTATGCTAAT	CTATGCTAAT	CTATGCTAAT		CTGATAAGAT	CTGATAAGAT	CTGATAAGAT	CTGATAAGAT			ATTAAACTCT	ATTAAACTCT	ATTAAACTCT	ATTAAACTCT			AACATTAGTA	AACATTAGTA	AACATTAGTA	AACATTAGTA	
601	NCAM c_1	NCAM c 2	$NCAM_d_2$	NCAM_d_1	651	NCAM C 1	NCAM C 2	NCAM d 2	NCAM_d_1		701	NCAM_c_1	NCAM_C_2	NCAM d 2	NCAM_d_1	<u>,</u>	TC/	NCAM_c_1	NCAM c_2	NCAM d 2	NCAM_d_1	

Fig. 1 (Cont.3)

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820	GAAtGGAGGA	GAAtGGAGGA	GAAtGGAGGA	GAAtGGAGGA	006	CTTaCAgcTG	CTTaCAgcTG	CTTACAGCTG	CTTACAGCTG	950	aCcaACATCA	accaACATCA	ACCAACATCA	ACCAACATCA	1000	AGATCCTTAT	AGATCCTTAT	AGATCCTtAT	AgATCCTtAT
	AGACTGTTTT	AGACTGTTTT	AGACTGTTTT	AGACTGTTTT		gatgcTGGTA	gatgcTGGTA	GATGCTGGTA	GATGCTGGTA		AAAAAAGTCC	AAAAAAGTCC	AAAAAAGTCC	AAAAAAGTCC		GGATCACACC	GGATCACACC	GGATCACACC	GGATCACACC
	CTGCCTGAAA	CTGCCTGAAA	CTGCCTGAAA	CTGCCTGAAA		CACCTcagat	CACCTcagat	CACCTCAGAT	CACCTCAGAT		GaAaCcCTGC	GaAaCcCTGC	GaAacccTGC	GaAaCCcTGC		GgaCGATTTT	GgaCGATTTT	GGaCGATTTT	GGaCGATTTT
	CTTTGGGACT	CTTTGGGACT	CTTTGGGACT	CTTTGGGACT		TACCTGCCAT	TACCTGCCAT	TACCIGCCAT	TACCIGCCAT		AATAATGTGG	AATAATGTGG	AATAATGTGG	AATAATGTGG		aTtAAAAAA	aTtAAAAAA	ATtAAAAAA	ATtAAAAAAA
	GGGTCAGGTC	GGGTCAGGTC	GGGTCAGGTC	GGGTCAGGTC		ACTTTGACCA	ACTTTGACCA	ACTTTGACCA	ACTTTGACCA		CATTGCCAAT	CATTGCCAAT	CATTGCCAAT	CATTGCCAAT		TTGtgAGAGC	TTGtgAGAGC	TTGTGAGAGC	TTGTGAGAGC
801				NCAM_d_1						901	NCAM c 1	NCAM c 2	NCAM d 2	NCAM d 1	951	NCAM c 1	NCAM c 2	NCAM d 2	NCAM_d_1

Fig. 1 (Cont. 4)

6/30

1050 TATCTTGCCA TATCTTGCCA tATCTTGCCA	1100 TTTAAAAATG TTTAAAAATG TTTAAAAATG	1150 GACTGATCCT GACTGATCCT GACTGATCCT GACTGATCCT	1200 TAAAATTCaC TAAAATTCaC TAAAATTCaC TAAAATTCaC
GAGGTGAAAA GAGGTGAAAA gAGGTGAAAA gAGGTGAAAA	ATTTAGTTGG ATTTAGTTGG aTTTAGTTGG aTTTAGTTGG	TCATTÀCACA TCATTACACA TCATTACACA TCATTACACA	ATCATTGATT ATCATTGATT ATCATTGATT
GATTGGCCGT GATTGGCCGT GATTGGCCGT	AGGAGCTAAC AGGAGCTAAC AGGAGCTAAC AGGAGCTAAC	GAGCGGATGG GAGCGGATGG GAGCGGATGG	AAACTTGGAC AAACTTGGAC AAACTTGGAC AAACTTGGAC
aCAACATCCA aCAACATCCA ACAACATCCA ACAACATCCA	GTTCCTTCTG GTTCCTTCTG GTTCCTTCTG	AAGAAGTTCT AAGAAGTTCT AAGAAGTTCT AAGAAGTTCT	CGGGAACAaC CGGGAACAaC CGGGAaCAaC CGGGAACAAC
CACAAAGATG CACAAAGATG CaCAAAGATG CaCAAAGATG	AGTAGAAGCT AGTAGAAGCT AGTAGAAGCT AGTAGAAGCT	GTCGTCCATT GTCGTCCATT GTCGTCCATT GTCGTCCATT	GATGTCTCTC GATGTCTCTC GATGTCTCTC GATGTCTCTC
1001 NCAM c 1 NCAM c 2 NCAM d 2 NCAM d 2	1051 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_1	1101 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_1	1151 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_1

Fig. 1 (Cont.⁵)

Fig. 1 (Cont.⁶)

7	1	3	X	0

1250	GgAGGaATAT	GGAGGAATAT	GGAGGAATAT	1300	TCCACCCAAT	TCCACCCAAT	TCCa	TCCa	1350	AAGGAGACAC	AAGGAGACAC	•	•	1400	ATCATCCTTT	ATCATCCTTT	•	•
	TCTGAAGGGA	TCTGAAGGGA	TCTGAAGGGA		GCAGCaCAGT	GCAGCACAGT	GCAGCACAGt	GCAGCACAGt		GTCACCAGAG	GTCACCAGAG	•	•		ACCTAAACCA	ACCTAAACCA	•	
	GTGTAGCATC	GTGTAGCATC	GTGTAGCATC		AATAtATCCA	AATAtATCCA	AATaTatCCA	AATaTatCCA		ATCACCATTG	ATCACCATTG	•	•		TAACTGGCAA	TAACTGGCAA	•	* * * * * * * * * * * * * * * * * * * *
		acGTACACAT ACGTACACAT	ACGTACACAT		TATCGATGTT	TATCGATGTT	TATCGATGTT	TATCGATGTT		CACAGGAAAA	CACAGGAAAA	•	•		CAATGTCAAG	CAATGTCAAG	•	•
	ggATTcTGGG	ggAITTCTGGG GGATTCTGGG	GGATTCTGGG		CTGATATCAG	CTGATATCAG	CTGATATCAG	CTGATATCAG		CIGACIGITC	CTGACTGTTC	•	•		AATAGAACTG	AATAGAACTG	•	
1201		NCAM c 2 NCAM d 2		1251	NCAM_c_1	NCAM_C_2	NCAM d 2	NCAM_d_1	1301	NCAM c 1	NCAM c 2	NCAM_d_2	NCAM_d_1	1351			NCAM_d_2	NCAM_d_1

1450 AATGCAAATG AATGCAAATG		1500	GGGAAATGTC GGGAAATGTC	•	•	1550	AACGTGAAAC	AACGTGAAAC	•	•	1600	TGCAGTGGAA	TGCAGTGGAA	TGCAGTGGAA	TGCAGTGGAA
CTGATGGATC		•	AATGTATCTA AATGTATCTA	•	•		CAATGGATTT	CAATGGATTT	•	•		AGTATCCCCC	AGTATCCCCC		
GTTGCAATGC		•	GAGGATTGTG GAGGATTGTG	•			CCAGCCAATA	CCAGCCAATA	•	•		CTCATCGTTC	CTCATCGTTC	•	•
GGATAAAGAA GGATAAAGAA		•	ATGGAACACT ATGGAACACT	•			AGATGTCAGa	AGATGTCAGa	•	•		CtIGGIGCAG	CtrgGrgCAG	•	•
GGTCTAGAGC	NCAM d 2	•	c_1 gAGAGTTATG AT	•			AGGAATGTAC	NCAM_c_2 AGGAATGTAC A	•	•		NCAM_c_1 CAAGGGAAGC	CAAGGGAAGC	•	•
1401 NCAM_c_1 NCAM_c_2	NCAM d 2	NCAM_a_1 1451	NCAM_c_1 NCAM_c_2	NCAM_d_2	NCAM_d_1	1501	NCAM_c_1	NCAM_C_2	NCAM d 2	NCAM a I	1551	NCAM c_1	NCAM c_2	NCAM_d_2	NCAM d 1

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1700 TATGAGTGGC TATGAGTGGC TATGAGTGGC	1750 TCAGGAATAC TCAGGAATAC TCAGGAATAC TCAGGAATAC	1800 GGGTTTATAA GGGTTTATAA GGGTTTATAA
GGTGCTGACC GGTGCTGACC GGTGCTGACC GGTGCTGACC	AATTTGACTC AATTTGACTC AALTTGACTC AALTTGACTC	GAAAACTATG GAAAACTATG GAAAACTATG GAAAACTATG
ATCCAATACG ATCCAATACG ATCCAATACG	CGGACGGGTC CGGACGGGTC CGGACGGGTC CGGACGGGTC	TCTTTCCAAT TCTTTCCAAT TCTTTCCAAT
CTGAGAGCCT CTGAGAGCCT CTGAGAGCCT CTGAGAGCCT	TAAATTATTA TAAATTATTA TAAATTATTA TAAATTATT	CTGTGAAGAG CTGTGAAGAG CTGTGAAGAG
TTGCAGAGTA TTGCAGAGTA TTGCAGAGTA TTGCAGAGTA	GCTTGGGCAA GCTTGGGCAA GCTTGGGCAA GCTTGGGCAA	ACAGAGTACG ACAGAGTACG ACAGAGTACG ACAGAGTACG
1651 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_2	1701 NCAM_c_1 NCAM_c_2 NCAM_d_2 NCAM_d_1	1751 NCAM c 1 NCAM c 2 NCAM d 2 NCAM d 1
	TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC	TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC TTGCAGAGTA CTGAGAGCCT ATCCAATACG GGTGCTGACC GCTTGGGCAA TAAATTATTA CGGACGGGTC AATTTGACTC GCTTGGGCAA TAAATTATTA CGGACGGGTC AATTTGACTC GCTTGGGCAA TAAATTATTA CGGACGGGTC AATTTGACTC GCTTGGGCAA TAAATTATTA CGGACGGGTC AATTTGACTC

Fig. 1(Cont.°)

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1850 TTTCTTGTTA TTTCTTGTTA TTTCTTGTTA	1900 CAATCCAGTA CAATCCAGTA CAATCCAGTA CAATCCAGTA	1950 GGACACAGAT GGACACAGAT GGACACAGAT	2000 GGCATCAGGC GGCATCAGGC GGCATCAGGC
GAGATGCAGC GAGATGCAGC GAGATGCAGC GAGATGCAGC	ATGATACCTA ATGATACCTA ATGATACCTA ATGATACCTA	AGTCTACAGT AGTCTACAGT AGTCTACAGT AGTCTACAGT	ATACCGGTTG ATACCGGTTG ATACCGGTTG
CTGGAGCTGG CTGGAGCTGG CTGGAGCTGG CTGGAGCTGG	GAATTCTATT GAATTCTATT GAATTCTATT GAATTCTATT	TTATTCTTAC TTATTCTTAC TTATTCTTAC	GGATTGTTGC GGATTGTTGC GGATTGTTGC GGATTGTTGC
ATAAATGAAG	CTATGCTCCA	GACACCGTGT	GCAGTGGATC
ATAAATGAAG	CTATGCTCCA	GACACCGTGT	GCAGTGGATC
ATAAATGAAG	CTATGCTCCA	GACACCGTGT	GCAGTGGATC
ATAAATGAAG	CTATGCTCCA	GACACCGTGT	GCAGTGGATC
CTGTAGCATC	CAGGAAAGGC	TGGCAGAACA	GAATCCTGAT
CTGTAGCATC	CAGGAAAGGC	TGGCAGAACA	GAATCCTGAT
CTGTAGCATC	CAGGAAAGGC	TGGCAGAACA	GAATCCTGAT
CTGTAGCATC	CAGGAAAGGC	TGGCAGAACA	GAATCCTGAT
1801	1851	1901	1951
NCAM c 1	NCAM c 1	NCAM c 1	NCAM c 1
NCAM c 2	NCAM c 2	NCAM c 2	NCAM c 2
NCAM d 2	NCAM d 2	NCAM d 2	NCAM d 2
NCAM d 1	NCAM d 2	NCAM d 2	NCAM d 2

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2050	IT AAATGGGAAT	I AAATGGGAAT	I Atalgggaar	T ALATGGGAAT	2100	C TAATTAAACC	C TAATTAAACC	C TAATTAAACC	C TAATTAAACC	2150	T GGTGAAGGAG	T GGTGAAGGAG	T GGTGAAGGAG	T GGTGAAGGAG	2200	A TCCTCATTTG	A TCCTCATTTG	A TCCTCATTTG	A TCCTCATTTG
	AGATTAAAAT	AGATTAAAAT	AGATTAAAAT	AGATTAAAAT		TTGACAGAGC	TTGACAGAGC	TTGACAGAGC	TTGACAGAGC		CACCAAATTT	CACCAAATTT	CACCAAATTT	CACCAAATTT		CTCCTGTAAA	CTCCTGTAAA	CTCCTGTAAA	CTCCTGTAAA
	TGGGAGCAGG	TGGGAGCAGG	TGGGAGCAGG	TGGGAGCAGG		GAGAATTAAT TACATATAAC	TACATATAAC	TACATATAAC	TACATATAAC	·	TGACTCCTCT	TGACTCCTCT	TGACTCCTCT	TGACTCCTCT		AAATATAGTG	AAATATAGTG	AAATATAGTG	AAATATAGTG
	GCAGCGCTGG	GCAGCGCTGG	GCAGCGCTGG	GCAGCGCTGG		GAGAATTAAT	GAGAATTAAT	GAGAATTAAT	GAGAATTAAT	•	GAAGTCCGAC	GAAGTCCGAC	GAAGTCCGAC	GAAGTCCGAC		TCGTGTGATC	TCGTGTGATC	TCGTGTGATC	TCGTGTGATC
	AGGCTGGACA	AGGCTGGACA	AGGCTGGACA	AGGCTGGACA		ATTCAAAAGG	ATTCAAAAGG	ATTCAAAAGG	ATTCAAAAGG		AGAAGCTTAT	AGAAGCTTAT	AGAAGCTTAT	AGAAGCTTAT		ATTCAACAAT	ATTCAACAAT	ATTCAACAAT	ATTCAACAAT
2001				NCAM_d_1						2101	$NCAM_{C_1}$	NCAM c 2	NCAM d 2	NCAM d 1	2151	NCAM_C_1			NCAM d 1

0677	TGTTCACTCA	TGTTCACTCA	TGTTCACTCA	TGTTCACTCA	2300	GCAACAAGAA	GCAACAAGAA	GCAACAAGAA	GCAACAAGAA	2350	TAGTGGCTCC	TAGTGGCTCC	TAGTGGCTCC	TAGTGGCTCC		2400	GATTGGaAGG	GATTGGaAGG	GATTGGaAGG	GATTGGaAGG
	AATATTTGTT	AATATTTGTT	AATATTTGTT	AATATTTGTT		GCAAAGTACA	GCAAAGTACA	GCAAAGTACA	GCAAAGTACA		ATGCTGACCG	ATGCTGACCG	ATGCTGACCG	ATGCTGACCG			TCACGACCCA	TCACGACCCA	TCACGACCCA	TCACGACCCA
	TGAAGATGGT	TGAAGATGGT	TGAAGATGGT	TGAAGATGGT		ACTGGACAAA	ACTGGACAAA	ACTGGACAAA	ACTGGACAAA		ACAGGACCTA	ACAGGACCTA	ACAGGACCTA	ACAGGACCTA			CATTGAGACA	CATTGAGACA	CATTGAGACA	CATTGAGACA
	ATTGTGGATT	ATTGTGGATT	ATTGTGGATT	ATTGTGGATT		GATAATTTG	GATAATTTTG	GATAATTTG	GATAATTTG		TACTCCTAAT	TACTCCTAAT	TACTCCTAAT	TACTCCTAAT			TTTATATGTA	TTTATATGTA	TTTATATGTA	TTTATATGTA
	AGAGAATTTC	AGAGAATTTC	AGAGAATTTC	AGAGAATTTC					AGATGATACA		ATACAAAATA	ATACAAAATA	ATACAAAATA	ATACAAAATA	•		AAAGAAGGTT	AAAGAAGGTT	AAAGAAGGTT	NCAM_d_1 AAAGAAGGTT 1
T022	NCAM c 1	$NCAM^{-2}$	NCAM d 2	NCAM_d_1 A	. 2251	NCAM c 1	$NCAM^{-2}$	NCAM d 2	NCAM_d_1	2301	$NCAM_c_1$	NCAM c 2	NCAM d 2	NCAM_d_1		2351	$NCAM_{c_1}$	NCAM c 2	NCAM_d_2	NCAM_d_1

Fig. $1(Cont.^{11})$

13/30

2450 CCCAAAAACC CCCAAAAACC CCCAAAAACC	2500 TTATCACATG TTATCACATG TTATCACATG	2550 TGAAAGGGCA TGAAAGGGCA TGAAAGGGCA TGAAAGGGCA	2600 AAAGGACAAA AAAGGACAAA AAAGGACAAA
CAGCATAGCT CAGCATAGCT CAGCATAGCT CAGCATAGCT	TCAGCTTCTT TCAGCTTCTT TCAGCTTCTT	TATCTACGTT TATCTACGTT TATCTACGTT TATCTACGTT	AAGTGGGAAT AAGTGGGAAT AAGTGGGAAT
GCCCTGTTTT GCCCTGTTTT GCCCTGTTTT	GCATATTGTT GCATATTGTT GCATATTGTT GCATATTGTT	CTTAAATGTT CTTAAATGTT CTTAAATGTT CTTAAATGTT	TGTGGTCTTC TGTGGTCTTC TGTGGTCTTC
CGACTTCTCA CGACTTCTCA CGACTTCTCA CGACTTCTCA	CaCAAACACT CaCAAACACT CaCAAACACT CaCAAACACT	ATATAGGTGT ATATAGGTGT ATATAGGTGT ATATAGGTGT	GAGAATCCAC GAGAATCCAC GAGAATCCAC GAGAATCCAC
CGAAAAGGCT CGAAAAGGCT CGAAAAGGCT CGAAAAGGCT	CTTATGGACC CTTATGGACC CTTATGGACC CTTATGGACC	TATGGACAAC TATGGACAAC TATGGACAAC TATGGACAAC	AACAACAATA AACAACAATA AACAACAATA AACAACAATA
2401 NCAM c 1 NCAM c 2 NCAM d 2 NCAM d 1	2451 NCAM_C_1 NCAM_C_2 NCAM_d_2 NCAM_d_1	2501 NCAM c 1 NCAM c 2 NCAM d 2 NCAM d 1	2551 NCAM c 1 NCAM c 2 NCAM d 2 NCAM d 2

Fig. 1 (Cont. 12)

TGATGGTGCT GTTGGGATTT Fig. 1 (Cont. 13)

2650	ATTTCAGCTC	ATTCAGCIC	ATTTCAGCTC	2700	TTGCTATTGA	TTGCTATTGA	TIGCIATIGA	TIGCTATIGA	2750	CTAGCAACTA	CTAGCAACTA	CTAGCAACTA	CTAGCAACTA	2800	ATGGCTTTTT	ATGGCTTTTT	ATGGCTTTTT	ATGGCTTTTT
	CAATTACTTC	CAATTACTIC	CAATTACTTC		GAAGGTGACA	GAAGGTGACA	GAAGGTGACA	GAAGGTGACA		AAAACAAGAC	AAAACAAGAC	AAAACAAGAC	AAAACAAGAC		TGGTTCATAT	TGGTTCATAT	TGGTTCATAT	TGGTTCATAT
	AATATATACC	AATATATACC	AATATATACC		TCCTGGAATA	TCCTGGAATA	TCCTGGAATA	TCCTGGAATA		GAGAATGTGC	GAGAATGTGC	GAGAATGTGC	GAGAATGTGC		GTTGGGATTT	GTTGGGATTT	GTTGGGATTT	GTTGGGATTT
	GGCTCATGTT		GGČTCATGTT		GTATCCGAGG	GTATCCGAGG	GTATCCGAGG	GTATCCGAGG		ATTGCAGAAG	ATTGCAGAAG	ATTGCAGAAG	ATTGCAGAAG		TGATGGTGCT	TGATGGTGCT	TGATGGTGCT	TGATGGTGCT
	GATGGAATGA	GATGGAATGA	GATGGAATGA		ATTTTCAAG	ATTTTGAAG	ATTTTCAAG	ATTTTCAAG		TGATGTATCA	TGATGTATCA	TGATGTATCA	TGATGTATCA		AGAATTCCGT	AGAATTCCGT	AGAATTCCGT	AGAATTCCGT
2601	NCAM C 1			2651	NCAM_c_1	NCAM c 2	NCAM d 2	NCAM_d_1	2701	NCAM_c_1	NCAM c 2	NCAM d 2	NCAM_d_1	2751	NCAM_c_1	NCAM_c_2	NCAM_d_2	NCAM_d_1

2850	GACCTTATCC	GACCTTATCC	GACCTTATCC	GACCTTATCC	2900	GAAAGAGTCT	GAAAGAGTCT	GAAAGAGTCT	GAAAGAGTCT		2950	TCCACTGACT	TCCACTGACT	TCCACTGACT	TCCACTGACT	3000	AGCACCTGGG	AGCACCTGGG	AGCACCTGGG	AGCACCTGGG
	CCTCGAAGGT	CCTCGAAGGT	CCTCGAAGGT	CCTCGAAGGT		CTGGCATGAA	CTGGCATGAA	CTGGCATGAA	CTGGCATGAA			CAAAGATTCC	CAAAGATTCC	CAAAGATTCC	CAAAGATTCC		AATTTTTTA	AATTTTTTA	AATTTTTTA	AATTTTTTA
	TATCTTAAGT	TATCTTAAGT	TATCTTAAGT	TATCTTAAGT		TCACCAGGCA	TCACCAGGCA	TCACCAGGCA	TCACCAGGCA			AACAAACTAC	AACAAACTAC	AACAAACTAC	AACAAACTAC		AATAAAAACA	AATAAAAACA	AATAAAAACA	AATAAAAACA
	TCCTCATCTC	TCCTCATCTC	TCCTCATCTC	TCCTCATCTC		TATAAAAGAT	TATAAAAGAT	TATAAAAGAT	TATAAAAGAT	٠		ACATTGAACA	ACATTGAACA	ACATTGAACA	ACATTGAACA		AAAATAAAT	AAAATAAAAT	AAAATAAAAT	AAAATAAAAT
	CCCATTATCG	CCCATTATCG	CCCATTATCG	CCCATTAICG		TGGCAGAGGC	TGGCAGAGGC	TGGCAGAGGC	TGGCAGAGGC			TTGTAAATGG	TTGTAAATGG	TTGTAAATGG	TTGTAAATGG		ACNGGACTCA	ACNGGACTCA	ACNGGACTCA	ACNGGACTCA
2801		$NCAM^{-c}$		NCAM_d_1	2851		2	NCAM d 2	NCAM_d_1		2901	NCAM c 1	0	NCAM d 2		2951	NCAM C 1	NCAM c 2	NCAM_d_2	NCAM d 1

AACATAAANG AACATAAANG AACATAAANG TINCAGACTA AACATAAANG 3050 TINCAGACTA TTNCAGACTA TINCAGACTA GTATAACTTA GTATAACTTA GTATAACTTA GTATAACTTA CATCATGGAA CATCATGGAA CATCATGGAA CATCATGGAA GATAAAAGA GATAAAAGA GATAAAAGA GATAAAAGA NCAM C 1 NCAM C 2 NCAM d 2 NCAM d 1

3051

NCAM_c_1 ATAATCGTTG ACCTG

NCAM_c_2 ATAATCGTTG ACCTG

NCAM_d_2 ATAATCGTTG ACCTG

NCAM_d_1 ATAATCGTTG ACCTG

Fig. 1 (Cont. 15)

Q	o l'I'l'CAGCATAGCTCCCAAAAATCCATATGGACCTACAAATAGTGCATATT 33	Ω.
2423	TITCAGCATAGCTCCCAAAACCCTTATGGACCCACAAACACTGCATATT	2472
56	GTTTCAGTTTCTATCACATGTACGGGCAACATATAGGGGGTTTTAAAT 1	105
2473		2522
106	GTATATCTACGTTTGAAAGGGCCAGACAACGATAGAGAATCCGCTATGGTC	155
2523	GTTTATCTACGTTTGAAAGGGCAAACAACAATAGAGAATCCACTGTGGTC	2572
156	GTCGAGTGGGAACAAGGACAACGATGGAATGAAGCTCATGTTAATATAT	205
2573	II	2622
.206	ATCCAATT.CTTCATTTCAGTTAATTTTTGAAGGCATTCGAGGTCCTGGG	254
. 2623	ACCCAATTACTTCATTTCAGCTCATTTTCGAAGGTATCCGAGGTCCTGGA 2672	2672

Fig.

	•		•					
304	354	2772	404	2822	454	2872	504	2922
ATAGAGGGTGACATCGCCATTGATGATGTATCAATTGCTGAAGGAGAATG	TGCAAAACAAGACCTACCAACTAAGAATTCCGTGGATGGTGCTGTTGGGA	TGCAAAACAAGACCTAGCAACTAAGAATTCCGTTGATGGTGCTGTTGGGA	TCTTAGTTCATATATGGCTTTTTCCAGTTATCATCTCATCTCTTA	- H	AGCCCTCGAAGGTGACCTTATCCTGGCAGAGGCTATTAATGATTCACCAG	AG	GCACTGGCATGAAGAAGAAGCTTTGTAAATGGACACTGAAAAAACAAAC	
255 2673	305	2723	355	2773	405	2823	455	2873

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Fig. 2 (Cont. 2)

4	FFINNABDONCHASAFFIANTAVISGEACHEERASENATITAETE S	2
 1	FFTNKAEDORCLASAPPTVRIVHSGLACNIEEERYSERVYTIREGETLEL 5	20
51	51 TCLVTGHPRPQIRWTKTAGSASDRFQDSSVFNETLRITNIQRHQGGRYYC 1	100
51		100
101	124	150
101	KAENGLGSPAIKSIRVDVYYLDDPVVTVHQSIGEAKEQFYYERTVFLRCV	150
151	ANSNPPVRYSWRRGQEVLLQGSDKGVEIYEPFFTQGETKILKLKNLRPQD	200
151		200
201	YANYSCIASVRNVCNIPDKMVSFRLSNKTASPSIKLLVDDPIVVNPGEAI	250
201		250

Fig.

Fig. 3 (Cont. 1)

251	TLVCVTTGGEPAPSLTWVRSFGTLPEKTVLNGGTLTIPAITSDDAGTYSC	300
251	TLVCVTTGGEPAPSLTWVRSFGTLPEKTVLNGGTLTIPALTSDDAGTYSC	300
301	IANNNVGNPAKKSTNIIVRALKKGRFWITPDPYHKDDNIQIGREVKISCQ	350
301	INNINGNPAKKSTNIIVRALKKGRFWITPDPYHKDDNIQIGREVKISCQ	350
351	VEAVPSEELTESWFKNGRPLRSSERMVITQTDPDVSPGTTNLDIIDLKFT	400
351	VEAVPSEELTESWEKNGRPLRSSERMVITQTDPDVSPGTTNLDIIDLKFT	400
401	DSGTYTCVASLKGGGISDISIDVNISSSTV	430
401	DSGTYTCVASLKGGGISDISIDVNISSSTVPPNLTVPQEKSPLVTREGDT	450
431		452
501	GMYRCQTSQYNGENVKPREALVQLIVQYPPAVEPAFLEIRQGQDRSVTMS	550

800	DDTDNFDWTKQSTATRNTKYTPNTGPNADRSGSKEGFYMYIETSRPRLEG	751
702	DDTDNEDWTKQSTATRNTKYTPNTGPNADRSGSKEGFYMYIETSRPRLEG	653
750	EAYEVRLTPLTKFGEGDSTIRVIKYSAPVNPHLREFHCGFEDGNICLFTQ	701
652	EAYEVRLTPLTKFGEGDSTIRVIKYSAPVNPHLREFHCGFEDGNICLFTQ	603
700	NPDAVDRIVAYRLGIRQAGQQRWWEQEIKINGNIQKGELITYNLTELIKP	. 651
602	NPDAVDRIVAYRLGIRQAGQQRWWEQEIKINGNIQKGELITYNLTELIKP	553
650		601
552	CSIINEAGAGRCSFLVTGKAYAPEFYYDTYNPVWQNRHRVYSYSLQWTQM	503
009	CRVLRAYPIRVLTYEWRLGNKLLRTGQFDSQEYTEYAVKSLSNENYGVYN	551
502	CRVLRAYPIRVLTYEWRLGNKLLRTGQFDSQEYTEYAVKSLSNENYGVYN	453

) i		
801	EKARLLSPVFSIAPKNPYGPTNTAYCFSFFYHMYGQHIGVLNVYLKLKGQ 850	
753		
851	TIENPLWSSSGNKGORWNEAHVNIYPITSFOLIFEGIRGPGIEGDIAID 900	
	•	
803	DVSIAEGECAKQDLATKNSVDGAVGILVHIWLFPIIVLISILSPRR*PYP 852	
(
901	DVSIAEGECAKODLATKNSVDGAVGILVHIWLFPIIVLISILSPRR*PYP 950	
α υ	COP DIHAGAMHATIANAIDEMIADIAA KAMAHDUKAAMHAMAAAADAD 858	
)))		
951	9	
	903 IKRHHGSITYXRLNIXDNR*P 923	

Fig. 3(Cont.3)

IKRHHGSITYXRLNIXDNR*P

	•	2-17-00	
TIREGETLEL ~~~~~~~~~ TIREGETLEL ~~~~~~~~~~	100 QRHQGGRYYC QRHQGGRYYC QRHQGGRYYC	150 YERTVFLRCV YERTVFLRCV YERTVFLRCV	200 LKLKNLRPQD LKLKNLRPQD LKLKNLRPQD LKLKNLRPQD
EEERYSERVY	FNETLRITNI FNETLRITNI FNETLRITNI FNETLRITNI	SIGEAKEQFY SIGEAKEQFY SIGEAKEQFY SIGEAKEQFY	PEFTQGETKI PEFTQGETKI PEFTQGETKI
IVHSGLACNI ~~~~~~~~~ IVHSGLACNI ~~~~~~~~~	ASDRFQDSSV ASDRFQDSSV ASDRFQDSSV ASDRFQDSSV	LDDPVVTVHQ LDDPVVTVHQ LDDPVVTVHQ LDDPVVTVHQ	GSDKGVEIYE GSDKGVEIYE GSDKGVEIYE GSDKGVEIYE
CLASAPPTVR	QIRWTKTAGS VHRWTKTAGS QIRWTKTAGS VHRWTKTAGS	IKSIRVDVYY IKSIRVDVYY IKSIRVDVYY IKSIRVDVYY	WRRGQEVLLQ WRRGQEVLLQ WRRGQEVLLQ WRRGQEVLLQ
FFTNKAEDQR ~~~~~~~~ FFTNKAEDQR ~~~~~~~~~	TCLVTGHPRP ~~~~~SQDIH TCLVTGHPRP ~~~~~SQDIH	KAENGLGSPA KAENGLGSPA KAENGLGSPA KAENGLGSPA	ANSNPPVRYS ANSNPPVRYS ANSNPPVRYS ANSNPPVRYS
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Fig. 4 (Cont. 1)

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4 (Cont. 4) Fig.

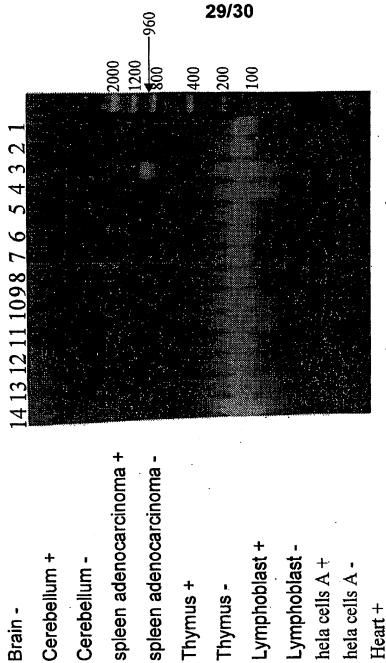
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ဖွဲ

Thymus +

Thymus -

ထ

Cerebellum +

Brain +

Brain -

Cerebellum -

+ superscript reverse transcriptase

superscript reverse transcriptase (control)

Fig. 5

10. Lymphoblast -

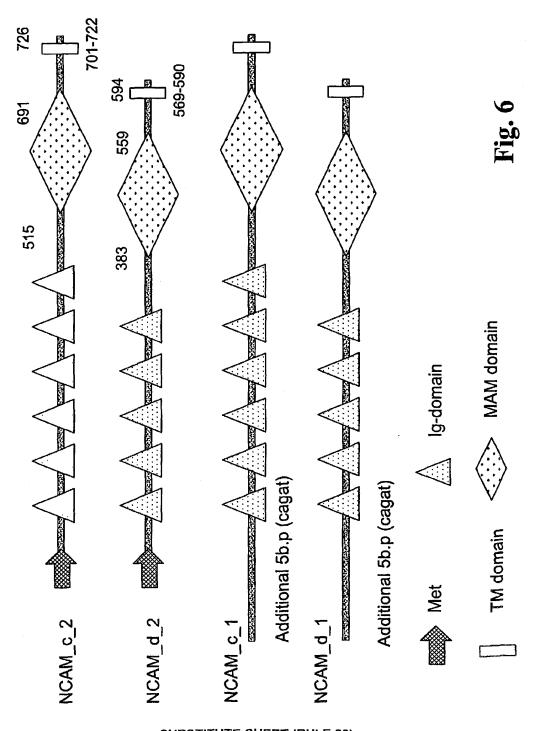
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SUBSTITUTE SHEET (RULE 26)

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Arg Tyr Tyr Cys Lys Ala Glu Asn Gly Leu Gly Ser Pro Ala Ile Lys
Ser Ile Arg Val Asp Val Tyr Tyr Leu Asp Asp Pro Val Val Thr Val
His Gln Ser Ile Gly Glu Ala Lys Glu Gln Phe Tyr Tyr Glu Arg Thr
Val Phe Leu Arg Cys Val Ala Asn Ser Asn Pro Pro Val Arg Tyr Ser
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Leu Lys Asn Leu Arg Pro Gln Asp Tyr Ala Asn Tyr Ser Cys Ile Ala
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Thr 465	Tyr	Glu	Trp	Arg	Leu 470	Gly	Asn	Lys	Leu	Leu 475	Arg	Thr	Gly	Gln	Phe 480
Asp	Ser	Gln	Glu	Tyr 485	Thr	Glu	Tyr	Ala	.Val 490	Lys	Ser	Leu	Ser	Asn 495	Glu
Asn	Tyr	Gly	Val 500	Tyr	Asn	Суз	Ser	Ile 505	Ile	Asn	Gļu	Ala	Gly 510	Ala	Gly
Arg	Cys	Ser 515	Phe	Leu	Val	Thr	Gly 520	Lys	Ala	Tyr	Ala	Pro 525	Glu	Phe	Tyr
Tyr	Asp 530	Thr	Tyr	Asn	Pro	Val 535	Trp	Gln	Asn	Arg	His 540	Arg	Val	Tyr	Ser
Tyr 545	Ser	Leu	Gln	Trp	Thr 550	Gln	Met	Asn	Pro	Asp 555	Ala	Val	Asp	Arg	Ile 560
Val	Ala	Tyr	Arg	Leu 565	Gly	Ile	Arg	Gln	Ala 570	Gly	Gln	Gln	Arg	Trp 575	Trp
Glu	Gln	Glu	Ile 580	Lys	Ile	Asn	Gly	Asn 585	Ile	Gln	Lys	Gly	Glu 590	Leu	Ile
Thr	Tyr	Asn 595	Leu	Thr	Glu	Leu	Ile 600	ъуs	Pro	Glu	Ala	Tyr 605	Glu	Val	Arg
Leu	Thr 610	Pro	Leu	Thr	Lys	Phe 615	Gly	Glu	Gly	Asp	Ser 620	Thr	Ile	Arg	Val
Ile 625	Lys	Tyr	Ser	Ala	Pro 630	Val	Asn	Pro	His	Leu 635	Arg	Glu	Phe	His	Cys 640
Gly	Phe	Glu	Asp	Gly 645	Asn	Ile	Cys	Lėu	Phe 650	Thr	Gln	Asp	Asp	Thr 655	Asp
Asn	Phe		Trp 660	Thr	Lys	Gln	Ser	Thr 665	Ala	Thr	Arg	Asn	Thr 670	Lys	Tyr
Thr	Pro	Asn 675	Thr	Gly	Pro	Asn	Ala 680	Asp	Arg	Ser	GIY	Ser 685	Lys	Glu	Gly
Phe	Tyr 690	Met	Tyr	Ile	Glu	Thr 695	Ser	Arg	Pro	Arg	Leu 700	Glu	GLy	Glu	Lys
Ala 705	Arg	Leu	Leu	Ser	Pro 710	Val	Phe	Ser	Ile	Ala 715	Pro	Lys ·	Asn	Pro	Tyr 720
Gly	Pro	Thr	Asn	Thr 725	Ala	Tyr	Cys	Phe	Ser 730	Phe	Phe	Tyr	His	Met 735	Tyr
Gly	Gln	His	Ile 740	Gly	Val	Leu	Asn	Val 745	Туг	Leu	Arg	Leu	Lys 750	Gly	Gln
Thr	Thr	Ile 755	Glu	Asn	Pro	Leu	Trp 760	Ser	Ser	Ser	Gly	Asn 765	Lys	Gly	Gln

Arg Trp Asn Glu Ala His Val Asn Ile Tyr Pro Ile Thr Ser Phe Gln 770 775 780

Leu Ile Phe Glu Gly Ile Arg Gly Pro Gly Ile Glu Gly Asp Ile Ala 785 790 795 800

Ile Asp Asp Val Ser Ile Ala Glu Glu Cys Ala Lys Gln Asp Leu 805 810

Ala Thr Lys Asn Ser Val Asp Gly Ala Val Gly Ile Leu Val His Ile 820 825 830

Trp Leu Phe Pro Ile Ile Val Leu Ile Ser Ile Leu Ser Pro Arg Arg 835 840 845

Pro Tyr Pro Gly Arg Gly Tyr Lys Arg Phe Thr Arg His Trp His Glu 850 860

Glu Arg Val Phe Val Asn Gly His Thr Asn Lys Leu Pro Lys Ile Pro 865 870 875 880

Pro Leu Thr Thr Gly Leu Lys Asn Lys Ile Ile Lys Thr Asn Phe Phe 885 890 895

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Glu Leu Thr Cys Leu Val Thr Gly His Pro Arg Pro Gln Ile Arg Trp
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Arg Tyr Tyr Cys Lys' Ala Glu Asn Gly Leu Gly Ser Pro Ala Ile Lys 100 105 110

Ser Ile Arg Val Asp Val Tyr Tyr Leu Asp Asp Pro Val Val Thr Val

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Val 145	Phe	Leu	Arg	Cys	Val 150	Ala	Asn	Ser	Asn	Pro 155	Pro	Val	Arg	Tyr	Ser 160
Trp	Arg	Arg	Gly	Gln 165	Glu	Val	Leu	Leu	Gln 170	Gly	Ser	Asp	Lys	Gly 175	Val
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Leu	Lys	Lys	Gly	Arg 325	Phe	Trp	Ile	Thr	Pro 330	Asp	Pro	Tyr	His	Lys 335	Asp
Asp	neA	Ile	Gln 340	Ile	Gly	Arg	Glu	Val 345	ГЛЗ	Ile	Ser	Cys	Gln 350	Val	Glu
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Ala	Val	Glu 435	Pro	Ala	Phe	Leu	Glu 440	Ile	Arg	Gln	Gly	Gln 445	Asp	Arg	Ser
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450 455 460 Thr Tyr Glu Trp Arg Leu Gly Asn Lys Leu Leu Arg Thr Gly Gln Phe " 475 Asp Ser Gln Glu Tyr Thr Glu Tyr Ala Val Lys Ser Leu Ser Asn Glu Asn Tyr Gly Val Tyr Asn Cys Ser Ile Ile Asn Glu Ala Gly Ala Gly Arg Cys Ser Phe Leu Val Thr Gly Lys Ala Tyr Ala Pro Glu Phe Tyr 520 Tyr Asp Thr Tyr Asn Pro Val Trp Gln Asn Arg His Arg Val Tyr Ser Tyr Ser Leu Gln Trp Thr Gln Met Asn Pro Asp Ala Val Asp Arg Ile Val Ala Tyr Arg Leu Gly Ile Arg Gln Ala Gly Gln Gln Arg Trp Trp Glu Gln Glu Ile Lys Ile Tyr Gly Asn Ile Gln Lys Gly Glu Leu Ile 585 Thr Tyr Asn Leu Thr Glu Leu Ile Lys Pro Glu Ala Tyr Glu Val Arg 600 Leu Thr Pro Leu Thr Lys Phe Gly Glu Gly Asp Ser Thr Ile Arg Val 615 The Lys Tyr Ser Ala Pro Val Asn Pro His Leu Arg Glu Phe His Cys Gly Phe Glu Asp Gly Asn I're Cys Leu Phe Thr Gln Asp Asp Thr Asp Asn Phe Asp Trp Thr Lys Gln Ser Thr Ala Thr Arg Asn Thr Lys Tyr 665 Thr Pro Asn Thr Gly Pro Asn Ala Asp Arg Ser Gly Ser Lys Glu Gly Phe Tyr Met Tyr Ile Glu Thr Ser Arg Pro Arg Leu Glu Gly Glu Lys 695 Ala Arg Leu Leu Ser Pro Val Phe Ser Ile Ala Pro Lys Asn Pro Tyr Gly Pro Thr Asn Thr Ala Tyr Cys Phe Ser Phe Phe Tyr His Met Tyr 730 Gly Gln His Ile Gly Val Leu Asn Val Tyr Leu Arg Leu Lys Gly Gln Thr Thr Ile Glu Asn Pro Leu Trp Ser Ser Ser Gly Asn Lys Gly Gln 760 Arg Trp Asn Glu Ala His Val Asn Ile Tyr Pro Ile Thr Ser Phe Gln 775

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Asn Gly Leu Gly Ser Pro Ala Ile Lys Ser Ile Arg Val Asp Val Tyr 50 60

Tyr Leu Asp Asp Pro Val Val Thr Val His Gln Ser Ile Gly Glu Ala 65 70 75 80

Lys Glu Gln Phe Tyr Tyr Glu Arg Thr Val Phe Leu Arg Cys Val Ala 85 90 95

Asn Ser Asn Pro Pro Val Arg Tyr Ser Trp Arg Arg Gly Gln Glu Val 100 105 110

Leu Leu Gln Gly Ser Asp Lys Gly Val Glu Ile Tyr Glu Pro Phe Phe 115 120 125

Thr Gln Gly Glu Thr Lys Ile Leu Lys Leu Lys Asn Leu Arg Pro Gln 130 135 140

Asp Tyr Aía Asn Tyr Ser Cys Ile Ala Ser Val Arg Asn Val Cys Asn 145 150 160

Ile Pro Asp Lys Met Val Ser Phe Arg Leu Ser Asn Lys Thr Ala Ser

Pro Ser Ile Lys Leu Leu Val Asp Asp Pro Ile Val Val Asn Pro Gly 180 185

Glu Ala Ile Thr Leu Val Cys Val Thr Thr Gly Gly Glu Pro Ala Pro 195 200 205

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Gl.u	Val 290	Lys	Ile	Ser	Cys	Gln 295	Val	Glu	Ala	Val	Pro 300	Ser	Glu	Glu	Leu
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345 350 Val Ala Ser Leu Lys Gly Gly Gly Ile Ser Asp Ile Ser Ile Asp Val 360 Asn Ile Ser Ser Ser Thr Val Pro Pro Asn Leu Thr Val Pro Gln Glu Lys Ser Pro Leu Val Thr Arg Glu Gly Asp Thr Ile Glu Leu Gln Cys 390 395 Gln Val Thr Gly Lys Pro Lys Pro Ile Ile Leu Trp Ser Arg Ala Asp Lys Glu Val Ala Met Pro Asp Gly Ser Met Gln Met Glu Ser Tyr Asp 425 Gly Thr Leu Arg Ile Val Asn Val Ser Arg Glu Met Ser Gly Met Tyr Arg Cys Gln Thr Ser Gln Tyr Asn Gly Phe Asn Val Lys Pro Arg Glu Ala Leu Val Gln Leu Ile Val Gln Tyr Pro Pro Ala Val Glu Pro Ala Phe Leu Glu Ile Arg Gln Gly Gln Asp Arg Ser Val Thr Met Ser Cys Arg Val Leu Arg Ala Tyr Pro Ile Arg Val Leu Thr Tyr Glu Trp Arg Leu Gly Asn Lys Leu Leu Arg Thr Gly Gln Phe Asp Ser Gln Glu Tyr 520 Thr Glu Tyr Ala Val Lys Ser Leu Ser Asn Glu Asn Tyr Gly Val Tyr 535 Asn Cys Ser Ile Ile Asn Glu Ala Gly Ala Gly Arg Cys Ser Phe Leu Val Thr Gly Lys Ala Tyr Ala Pro Glu Phe Tyr Tyr Asp Thr Tyr Asn Pro Val Trp Gln Asn Arg His Arg Val Tyr Ser Tyr Ser Leu Gln Trp 585 Thr Gln Met Asn Pro Asp Ala Val Asp Arg Ile Val Ala Tyr Arg Leu 600 Gly Ile Arg Gln Ala Gly Gln Gln Arg Trp Trp Glu Gln Glu Ile Lys 615 Ile Asn Gly Asn Ile Gln Lys Gly Glu Leu Ile Thr Tyr Asn Leu Thr Glu Leu Ile Lys Pro Glu Ala Tyr Glu Val Arg Leu Thr Pro Leu Thr Lys Phe Gly Glu Gly Asp Ser Thr Ile Arg Val Ile Lys Tyr Ser Ala 665

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Leu	Lys	Lys	Gly	Arg 325	Phe	Trp	Ile	Thr	Pro 330	Asp	Pro	Tyr	His	Lys 335	Asp
Asp	Asn	Ile	Gln 340	Ile	Gly	Arg	Glu	Val 345	Lys	Ile	Ser	Cys	Gln 350	Val	Glu
Ala	Val	Pro 355	Ser	Glu	Glu	Leu	Thr 360	Phe	Ser	Trp	Phe	Lys 365	Asn	Gly	Arg

Val Ser Pro Gly Thr Thr Asn Leu Asp Ile Ile Asp Leu Lys Phe Thr Asp Ser Gly Thr Tyr Thr Cys Val Ala Ser Leu Lys Gly Gly Gly Ile Ser Asp Ile Ser Ile Asp Val Asn Ile Ser Ser Ser Thr Val Pro Pro Asn Leu Thr Val Pro Gln Glu Lys Ser Pro Leu Val Thr Arg Glu Gly Asp Thr Ile Glu Leu Gln Cys Gln Val Thr Gly Lys Pro Lys Pro Ile Ile Leu Trp Ser Arg Ala Asp Lys Glu Val Ala Met Pro Asp Gly Ser Met Gln Met Glu Ser Tyr Asp Gly Thr Leu Arg Ile Val Asn Val Ser Arg Glu Met Ser Gly Met Tyr Arg Cys Gln Thr Ser Gln Tyr Asn Gly 505 Phe Asn Val Lys Pro Arg Glu Ala Leu Val Gln Leu Ile Val Gln Tyr Pro Pro Ala Val Glu Pro Ala Phe Leu Glu Ile Arg Gln Gly Gln Asp Arg Ser Val Thr Met Ser Cys Arg Val Leu Arg Ala Tyr Pro Ile Arg Val Leu Thr Tyr Glu Trp Arg Leu Gly Asn Lys Leu Leu Arg Thr Gly Gln Phe Asp Ser Gln Glu Tyr Thr Glu Tyr Ala Val Lys Ser Leu Ser Asn Glu Asn Tyr Gly Val Tyr Asn Cys Ser Ile Ile Asn Glu Ala Gly Ala Gly Arg Cys Ser Phe Leu Val Thr Gly Lys Ala Tyr Ala Pro Glu Phe Tyr Tyr Asp Thr Tyr Asn Pro Val Trp Gln Asn Arg His Arg Val 635 Tyr Ser Tyr Ser Leu Gln Trp Thr Gln Met Asn Pro Asp Ala Val Asp 650 Arg Ile Val Ala Tyr Arg Leu Gly Ile Arg Gln Ala Gly Gln Gln Arg Trp Trp Glu Gln Glu Ile Lys Ile Asn Gly Asn Ile Gln Lys Gly Glu 680 Leu Ile Thr Tyr Asn Leu Thr Glu Leu Ile Lys Pro Glu Ala Tyr Glu Val Arg Leu Thr Pro Leu Thr Lys Phe Gly Glu Gly Asp Ser Thr Ile

705					710					715					720
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Thr	Asp	Asn 755	Phe	Asp	Trp	Thr	Lys 760	Gln	Ser	Thr	Ala	Thr 765	Arg	Asn	Thr
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Glu 785	Gly	Phe	Tyr	Met	Tyr 790	Ile	Glu	Thr	Ser	Arg 795	Pro	Arg	Leu	Glu	Gly 800
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Asp	Leu	Ala 915	Thr	Lys	Asn	Ser	Val 920	Asp	Gly	Ala	Val	Gly 925	Ile	Leu	Val
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Arg 945	Arg														•

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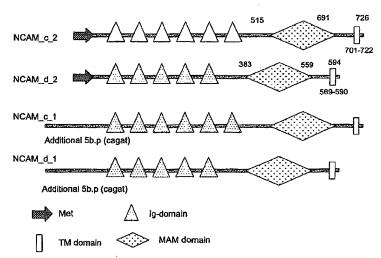
with international search report

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27 December 2001

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: N-CAM HOMOLOG AND SPLICE VARIANTS THEREOF



(57) Abstract: The invention concerns several nucleic acid and amino acid sequences which are homologs of the neuronal cell adhesion molecule. These sequences are splice variants of each other.



INTERNATIONAL SEARCH REPORT

In' ational Application No PCT/IL 00/00664

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A. CLASSI IPC 7	FICATION OF SUBJECT MATTER C12N15/12 C07K14/705 C07K16/A61K39/395 C12Q1/68 G01N33/		17 A61I	K31/70
According to	o International Patent Classification (IPC) or to both national classific	cation and IPC		
B. FIELDS	SEARCHED			
Minimum do IPC 7	ocumentation searched (classification system followed by classification C12N C07K A61K C12Q G01N	tion symbols)		
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Electronic d	ata base consulted during the international search (name of data b	ase and, where practical,	search terms use	d)
	ENTS CONSIDERED TO BE RELEVANT		· · · · · · · · · · · · · · · · · · ·	
Category °	Citation of document, with indication, where appropriate, of the re	elevant passages		Relevant to claim No.
X	DATABASE EMBL 'Online! Acc.no. AI859192, 22 July 1999 (1999-07-22) STRAUSBERG, R. ET AL.: "w166h10. NCI_CGAP_Brn25 Homo sapiens cDNA IMAGE:2429923 3' similar to SW:TI Q91641 THYROID HORMONE-INDUCED P PRECURSOR." XP002173300 Human EST with 99.1% identity wi seq.ID.1 between nt.2516-2961. the whole document	clone HIB_XENLA ROTEIN B		1-4
X Furth	er documents are listed in the continuation of box C.	X Patent family m	nembers are listed	in annex.
"A" docume conside "E" earlier diffing di "L" documer which i citation "O" docume other n	nt which may throw doubts on priority claim(s) or s cited to establish the publication date of another or other special reason (as specified) nt referring to an oral disclosure, use, exhibition or leans nt published prior to the international filing date but	cited to understand invention "X" document of particul cannot be consider involve an inventive "Y" document of particul cannot be consider document is combinent, such combinent in the art.	not in conflict with the principle or the ar relevance; the ced novel or cannol or step when the do ar relevance; the ced to involve an in- ned with one or mo- nation being obvious	the application but early underlying the claimed invention to considered to comment is taken alone claimed invention ventive step when the one other such docuust o a person skilled
	an the priority date claimed	*4. document member o		
	7 July 2001	07/08/20		
Name and m	ailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Smalt, R		

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INTERNATIONAL SEARCH REPORT

Ir 'ational Application No
PCT/IL 00/00664

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	ation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the rokevant passages		Relevant to claim No.
A	SAUGIER-VEBER PASCALE ET AL: "Identification of novel L1CAM mutations using fluorescence-assisted mismatch analysis." HUMAN MUTATION, vol. 12, no. 4, 1998, pages 259-266, XP000990828 ISSN: 1059-7794 the whole document		
A	US 5 840 689 A (DANILOFF JOANNE K) 24 November 1998 (1998-11-24) the whole document		
A	WO 96 04396 A (SYSTEMIX INC) 15 February 1996 (1996-02-15) the whole document		
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FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Present claims 22 and 23 relate to an activator and a deactivator, respectively, of the protein of the invention.

The application provides no support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for any such compounds. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the compounds by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, the search has been carried out for those parts of the claims which appear to be clear, namely those parts relating to the (de)activators which can be defined by the protein of the invention, namely antibodies directed to said protein.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Ir' national Application No

	Information on patent family members				Puf/IL 00/00664	
Patent document cited in search report		Publication date	Patent family member(s)		Publication date	
US 5840689	Α	24-11-1998	NONE			
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